

Subsystems Annotation: the Key to Consistent, Accurate Useful Annotations

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What Should be the Goal of the Annotation Effort?

- Consistent, accurate annotations of function
- Grouped into operational subcomponents, which we call “subsystems”
- Establish internal consistency needed to reconstruct the metabolic network
- Do this in a way that scales

Example Subsystem: Histidine Degradation

- Conversion of histidine to glutamate
- Functional roles defined in table
- Inclusion in subsystem is *only* by functional role
- Controlled vocabulary ...

Subsystem: Histidine Degradation		
1	HutH	Histidine ammonia-lyase (EC 4.3.1.3)
2	HutU	Urocanate hydratase (EC 4.2.1.49)
3	HutI	Imidazolonepropionase (EC 3.5.2.7)
4	GluF	Glutamate formiminotransferase (EC 2.1.2.5)
5	HutG	Formiminoglutamase (EC 3.5.3.8)
6	NfoD	N-formylglutamate deformylase (EC 3.5.1.68)
7	ForI	Formiminoglutamic iminohydrolase (EC 3.5.3.13)

Subsystem Spreadsheet

Subsystem Spreadsheet									
Organism	Variant	HutH	HutU	HutI	GluF	HutG	NfoD	ForI	
<i>Bacteroides thetaiotaomicron</i>	1	Q8A4B3	Q8A4A9	Q8A4B1	Q8A4B0				
<i>Desulfotela psychrophila</i>	1	gi 51246205	gi 51246204	gi 51246203	gi 51246202				
<i>Halobacterium sp.</i>	2	Q9HOD5	Q9HOD8	Q9HOD6		Q9HOD7			
<i>Deinococcus radiodurans</i>	2	Q9RZ06	Q9RZ02	Q9RZ05		Q9RZ04			
<i>Bacillus subtilis</i>	2	P10944	P25593	P42084		P42068			
<i>Caulobacter crescentus</i>	3	P58682	Q9A9M1	P58079			Q9A9M0	Q9A9L9	
<i>Pseudomonas putida</i>	3	Q88CZ7	Q88CZ6	Q88CZ9			Q88D00	Q88CZ3	
<i>Xanthomonas campestris</i>	3	Q8PAA7	P58988	Q8PAA6			Q8PAA8	Q8PAA5	
<i>Listeria monocytogenes</i>	-1								

- Column headers taken from table of functional roles
- Rows are selected genomes or organisms
- Cells are populated with specific, annotated genes
- Functional variants defined by the annotated roles
- Variant code -1 indicates subsystem is not functional
- Clustering shown by color

“The Populated Subsystem”

Subsystem: Histidine Degradation	
1	HutH
2	HutU
3	HutI
4	GluF
5	HutG
6	NfoD
7	ForI
	Histidine ammonia-lyase (EC 4.3.1.3)
	Urocanate hydratase (EC 4.2.1.49)
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Subsystem Spreadsheet									
Organism	Variant	HutH	HutU	HutI	GluF	HutG	NfoD	ForI	
<i>Bacteroides thetaiotomicron</i>	1	Q8A4B3	Q8A4A9	Q8A4B1	Q8A4B0				
<i>Desulfovibrio psychrophila</i>	1	gi 51246205	gi 51246204	gi 51246203	gi 51246202				
<i>Halobacterium sp.</i>	2	Q9HOD5	Q9HOD8	Q9HQD6		Q9HOD7			
<i>Deinococcus radiodurans</i>	2	Q9RZ06	Q9RZ02	Q9RZ05		Q9RZ04			
<i>Bacillus subtilis</i>	2	P10944	P25503	P42084		P42068			
<i>Caulobacter crescentus</i>	3	PS8082	Q9A9M1	PS8079			Q9A9M0	Q9A9L9	
<i>Pseudomonas putida</i>	3	Q8RCZ7	Q8RCZ6	Q8RCZ9			Q8SD00	Q8SCZ3	
<i>Xanthomonas campestris</i>	3	Q8PA47	PS8988	Q8PA46			Q8PA48	Q8PA45	
<i>Listeria monocytogenes</i>	-1								

Subsystems Can Be Made as “Stand-Alone” Data Objects

- This only requires conversion of gene/protein IDs
- It establishes a consistent, controlled vocabulary
- You can annotate the functional roles (with reactions, GO terms, literature) and achieve propagation to genes

Increased Accuracy Involves Quality Control and Consistency Checks

- New wet lab data must be integrated rapidly
- Inconsistencies in the metabolic reconstruction must be revealed, maintained, and (eventually) corrected
- Focusing literature on functional roles is essential to accurate propagation
- Connecting the implications of assignments to phenotypic measurements is essential for quality control

The Problem:

- Develop subsystems that cover what is known about the genes/proteins
- Maintain them (with experimental data)
- Develop high-throughput tools to integrate new genomes into the spreadsheet
- Construct the induced protein families