

Model SEED Tutorial Part 1: Technical Primer

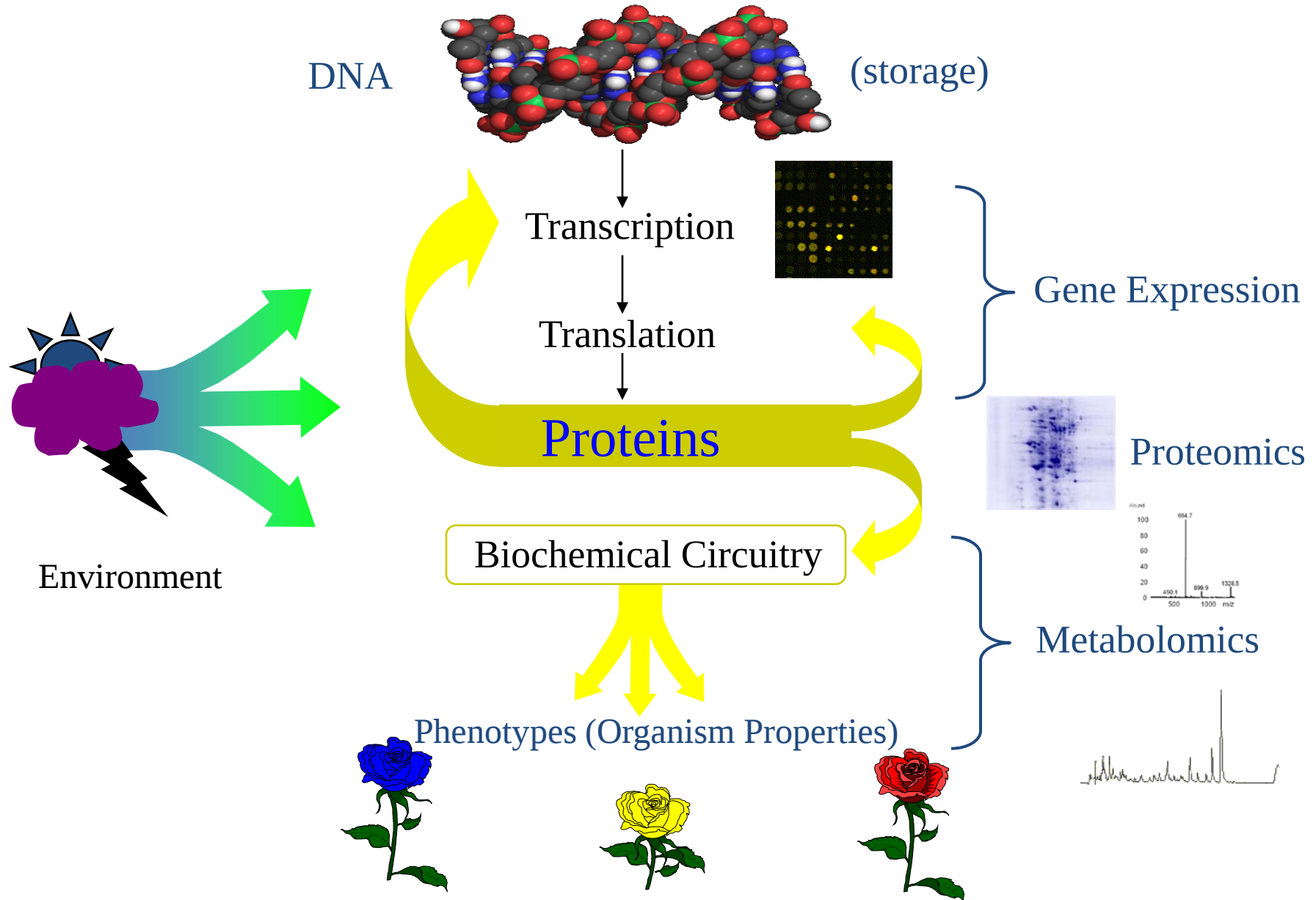
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Presented by: Christopher Henry

August 31-September 2



***Predicting Phenotypes from Genotypes* —the prediction of system level behavior from collections of functional components**



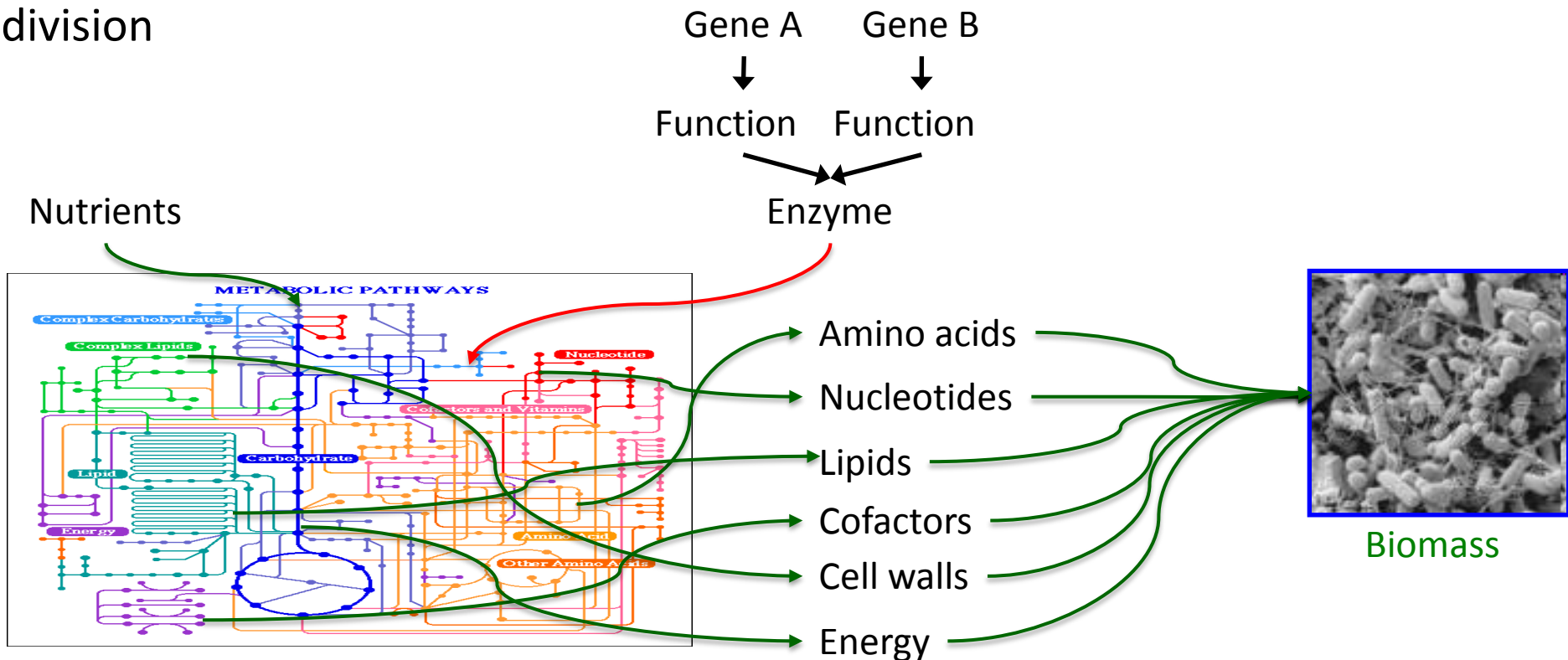
Adapted From Bruno Sobral VBI

Genome + Environment = Phenotype

Metabolic Modeling is One Key to Predicting Phenotype from Genotype

What is a metabolic model?

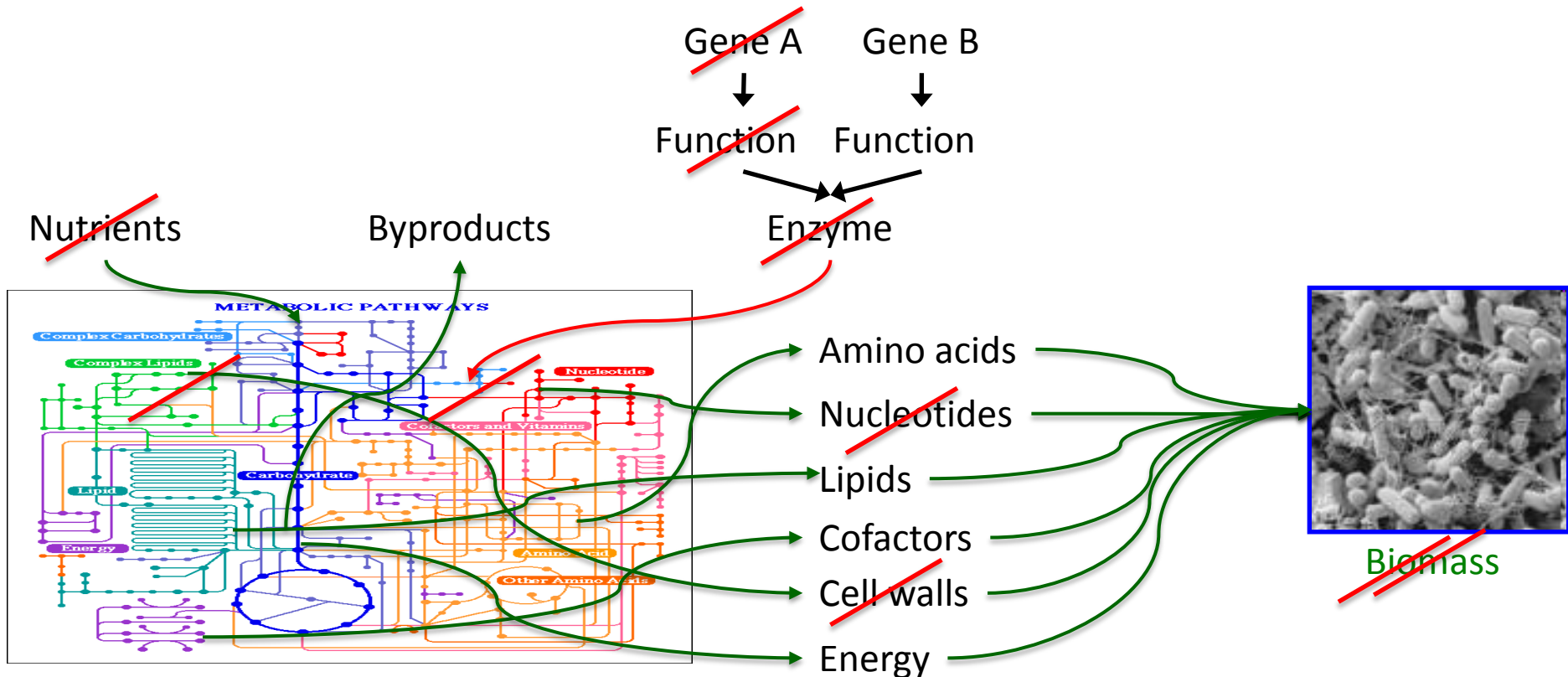
- 1.) A list of all reactions involved in the metabolic pathways
- 2.) A list of rules associating reaction activity to gene activity
- 3.) A biomass reaction listing essential building blocks needed for growth and division



Metabolic Modeling is One Key to Predicting Phenotype from Genotype

What can a metabolic model do?

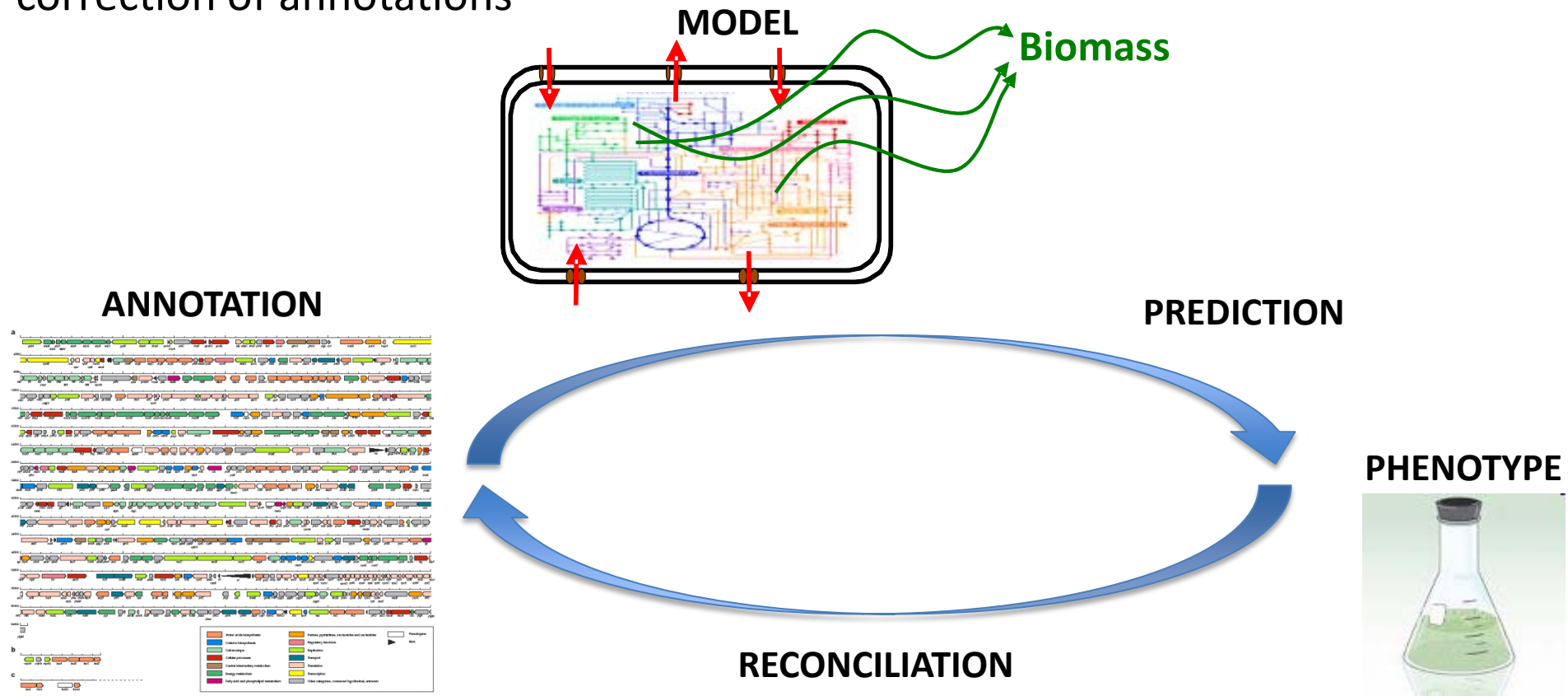
- 1.) Predict culture conditions and possible responses to environment changes.
- 2.) Predict metabolic capabilities from genotype.
- 3.) Predict impact of genetic perturbations



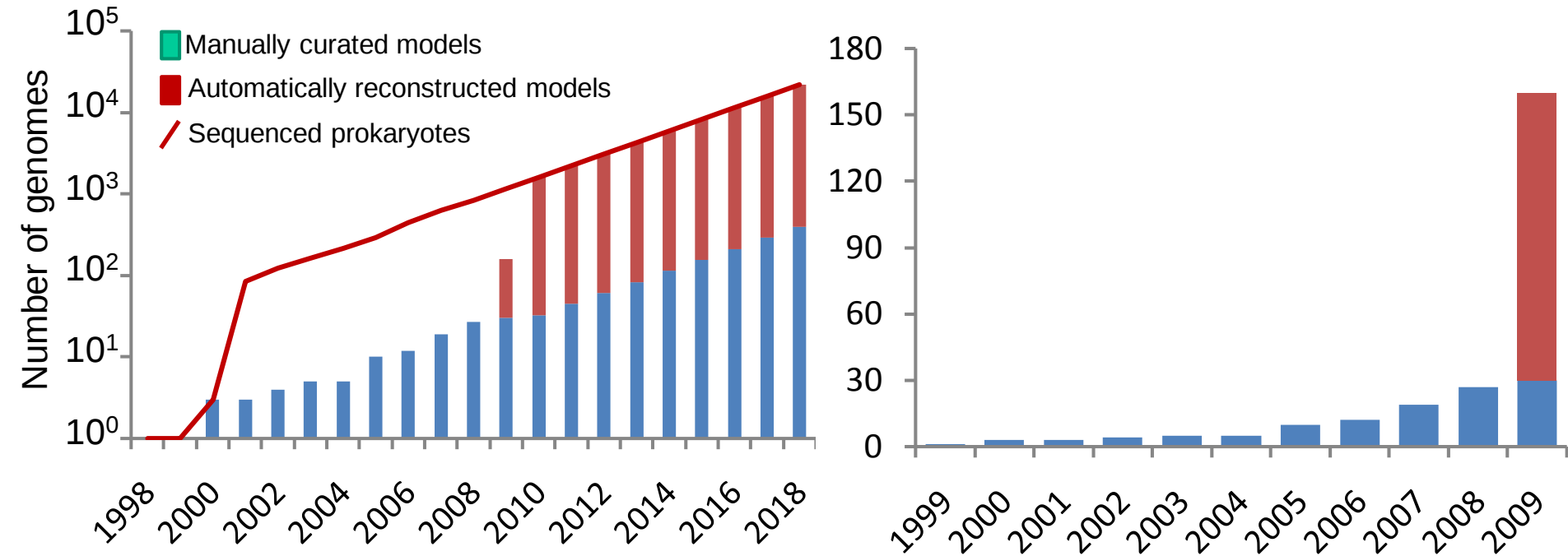
Metabolic Modeling is One Key to Predicting Phenotype from Genotype

What can a metabolic model do?

- 1.) Predict culture conditions and possible responses to environment changes.
- 2.) Predict metabolic capabilities from genotype.
- 3.) Predict impact of genetic perturbations
- 4.) Linking annotations to observed organism behavior enabling validation and correction of annotations

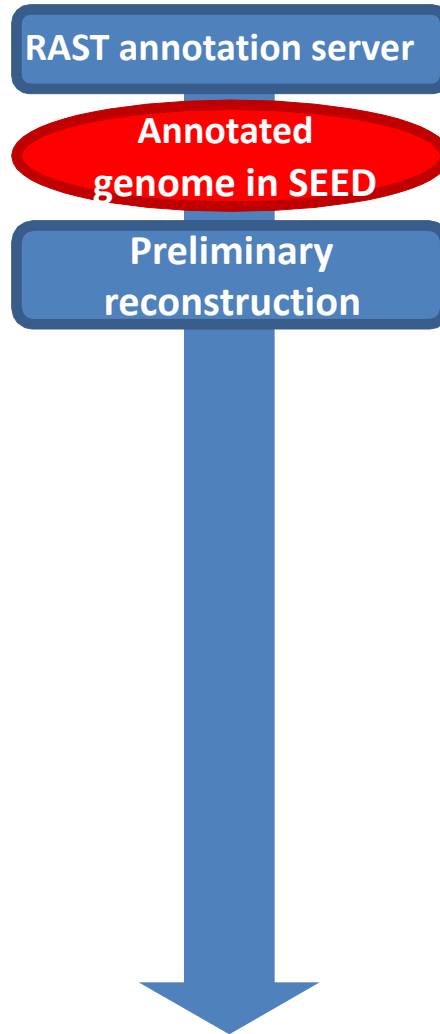


Model reconstruction lags behind genome sequencing



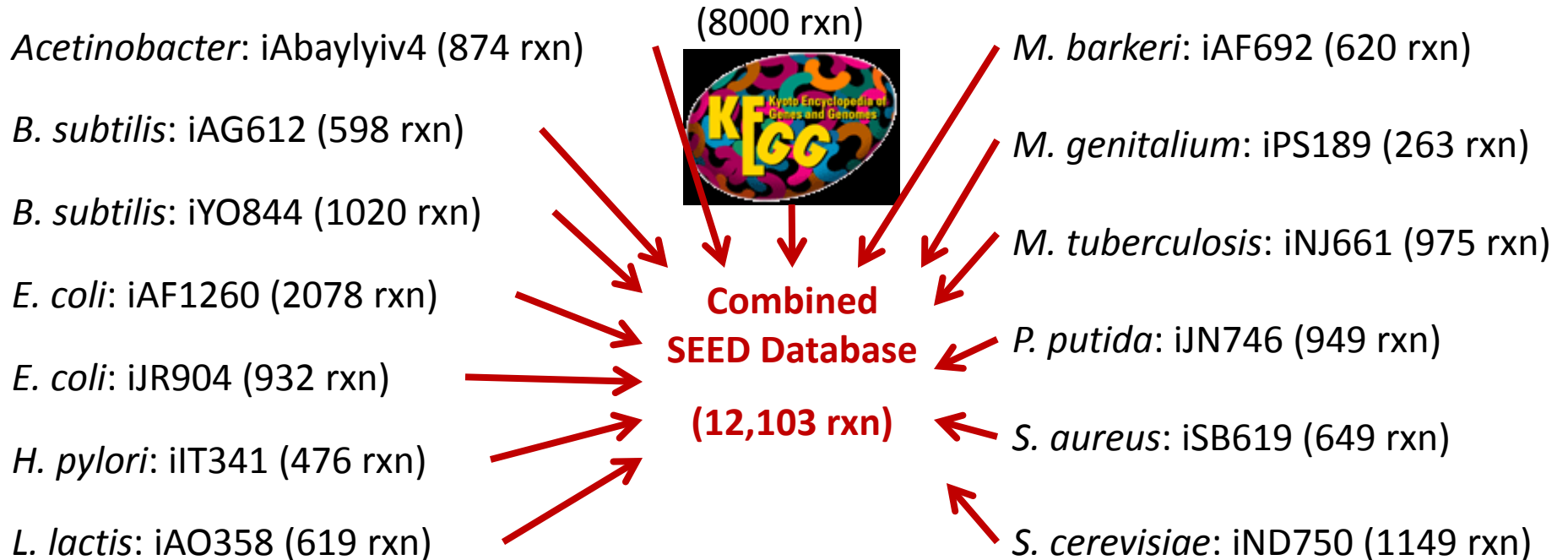
- **≈1000** completely sequenced prokaryotes vs **≈30** published genome-scale models
- Models are often constructed one-at-a-time by individuals working independently
- Model building typically begins by identifying bidirectional best hits with *E. coli*
- Current process results in replication of work, propagation of errors, and extensive manual curation
- Bottom line: it previously required approximately one year to produce a complete model

Model SEED: Converting Annotated Genomes into Genome-scale Metabolic Models



Biochemistry Database in the SEED

- A biochemistry database was constructed combining content from the **KEGG** and **13** published genome-scale models into a non-redundant set of compounds and reactions

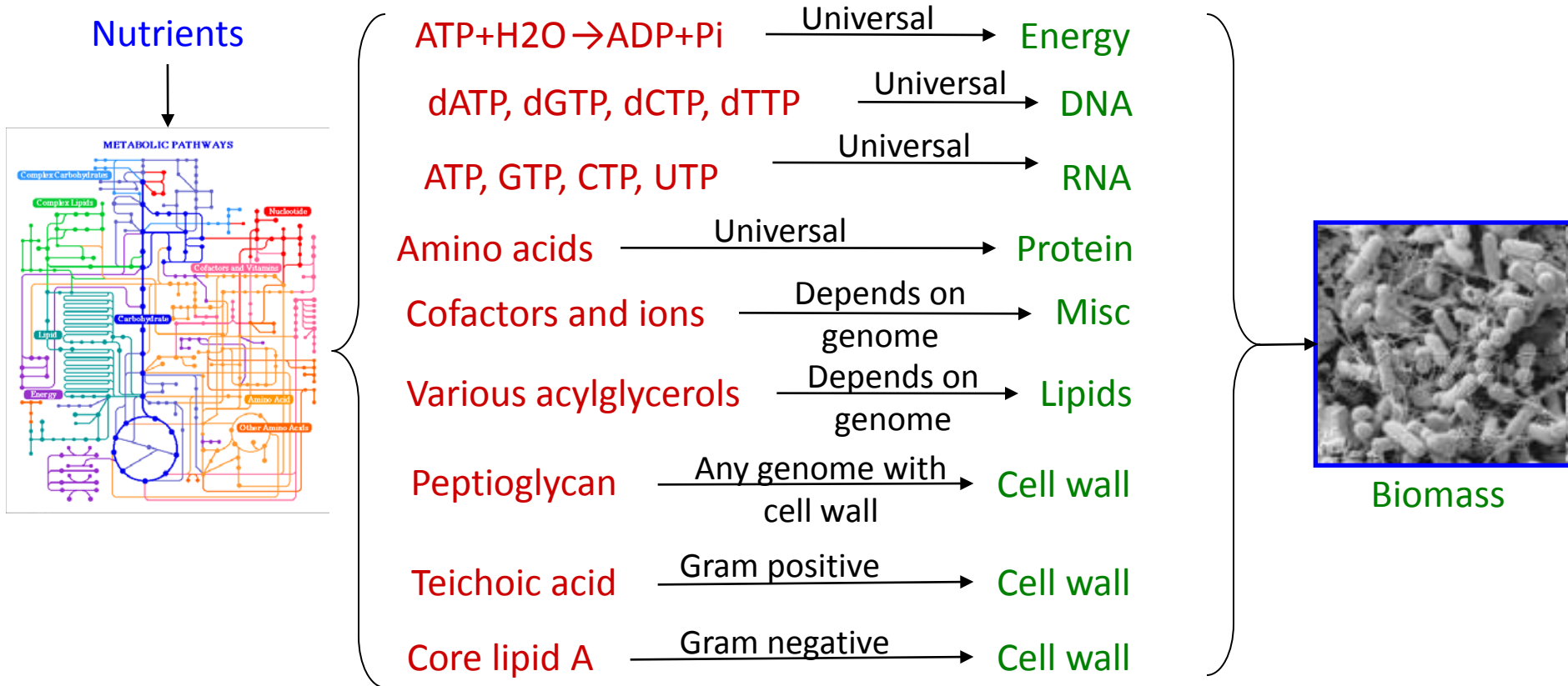


- Reactions were then mapped to the functional roles in the SEED based on EC number, substrate names, and enzyme names:

REACTION	COMPLEX	FUNCTIONAL ROLE	GENE
$\text{NAD}^+ + \text{NADPH} \leftrightarrow \text{NADH} + \text{NADP}^+$	→ Gene complex	NAD(P) transhydrogenase subunit beta (EC 1.6.1.2)	→ peg.100
		NAD(P) transhydrogenase alpha subunit (EC 1.6.1.2)	→ peg.101

Biomass Objective Function

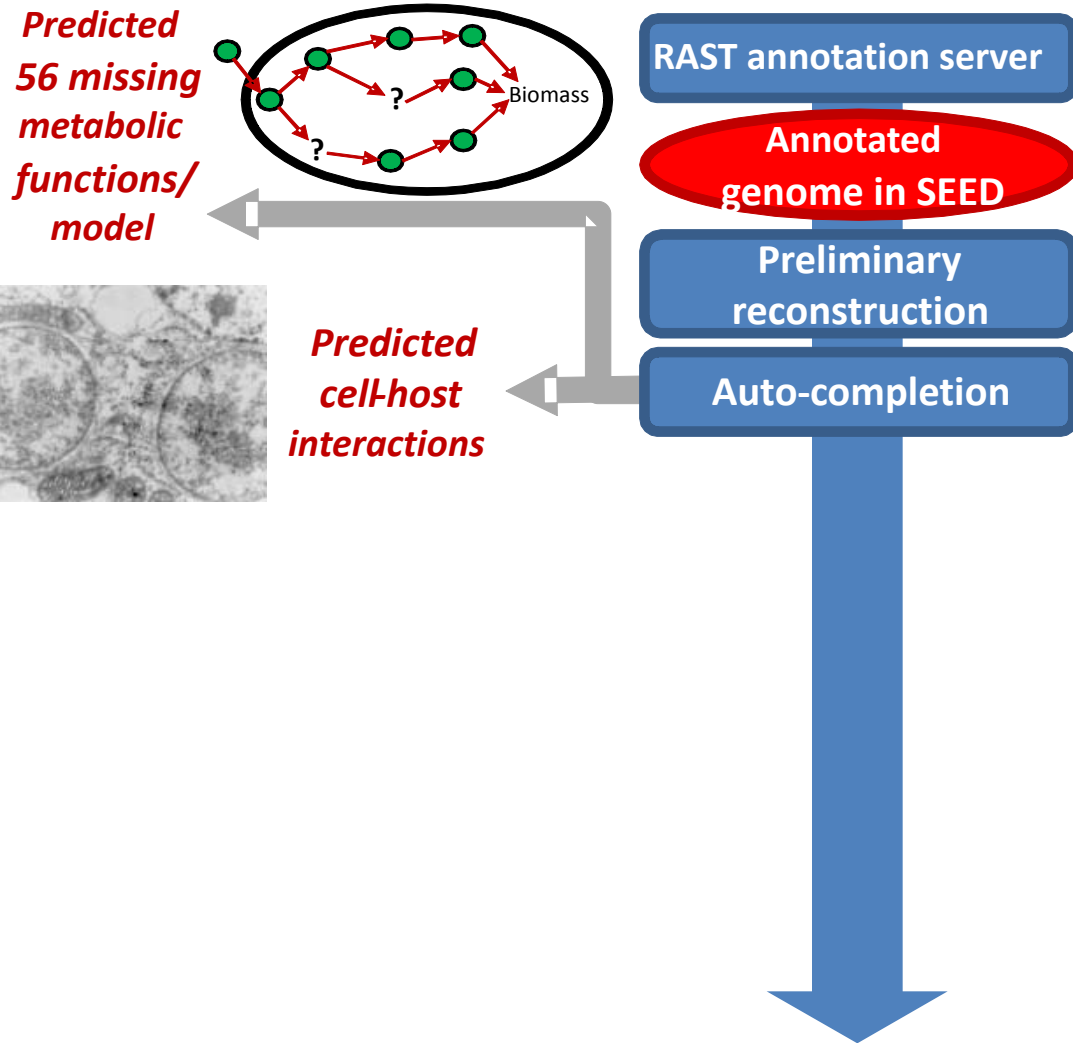
- To test growth of the model, we build a biomass objective function template



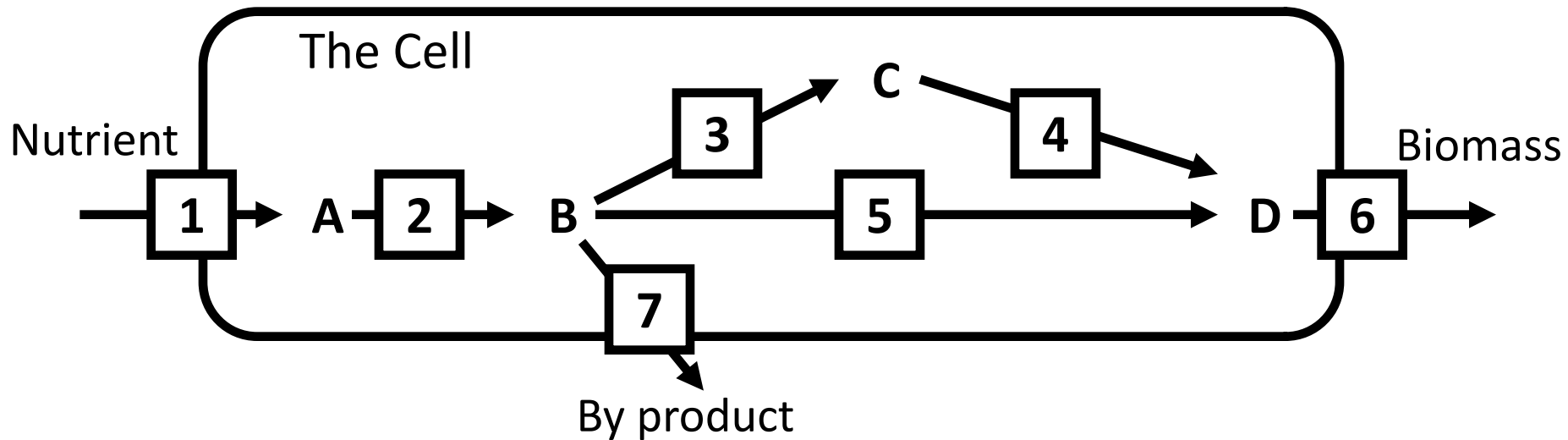
- Each biomass component may be rejected from the biomass reaction of a model based on the following criteria:

- Subsystem representation
- Functional role presence
- Taxonomy
- Cell wall types

Model SEED: Converting Annotated Genomes into Genome-scale Metabolic Models



Flux Balance Analysis



Assuming Steady State:

No internal metabolite is
allowed to accumulate

Thus, reaction rates are constrained
by mass balances

For example:

At Steady State:

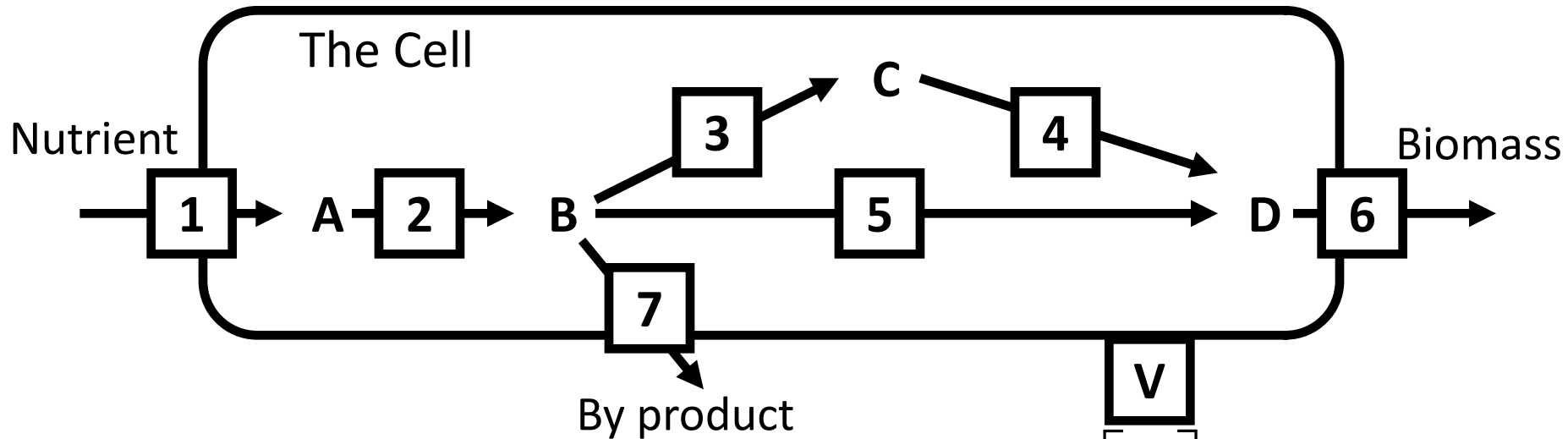
$$v_1 = v_2$$

$$v_2 = v_3 + v_5 + v_7$$

$$v_3 = v_4$$

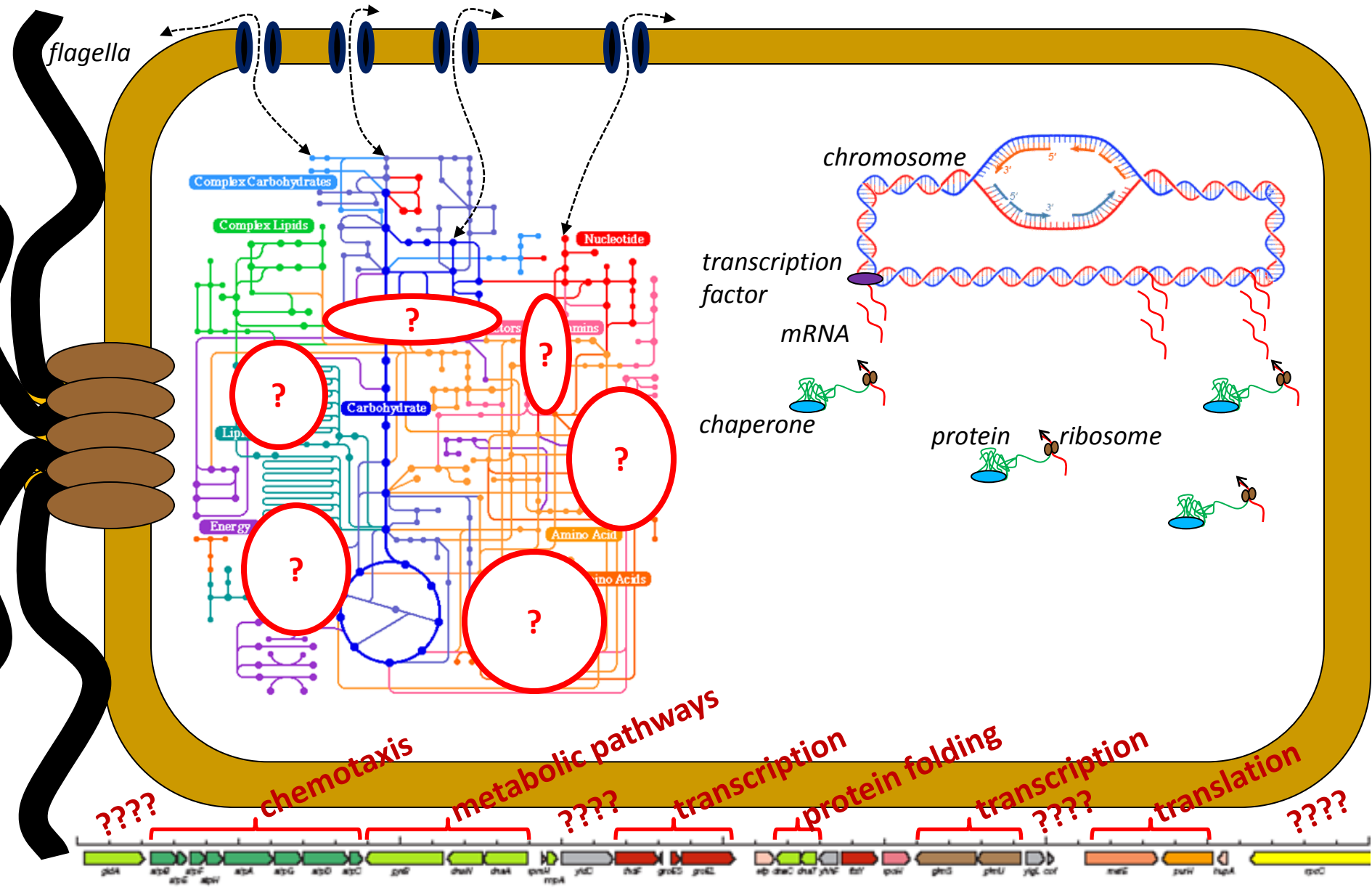
$$v_4 + v_5 = v_6$$

Flux Balance Analysis

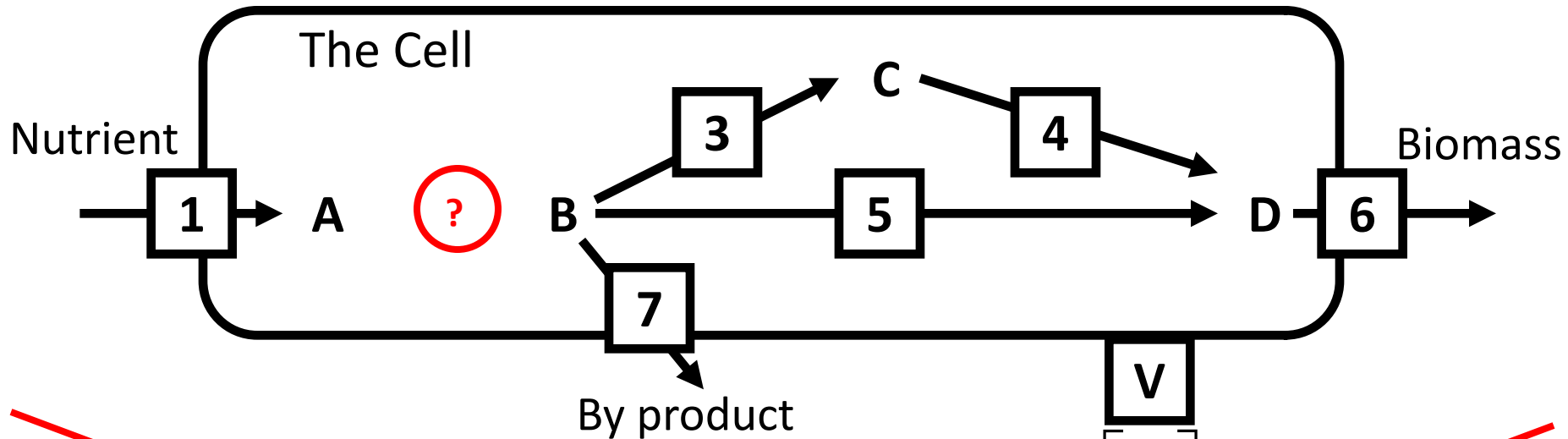


$$\begin{array}{c}
 \begin{array}{c} \mathbf{A} \\ \mathbf{B} \\ \mathbf{C} \\ \mathbf{D} \end{array}
 \begin{bmatrix}
 \boxed{1} & \boxed{2} & \boxed{3} & \boxed{4} & \boxed{5} & \boxed{6} & \boxed{7} \\
 1 & -1 & 0 & 0 & 0 & 0 & 0 \\
 0 & 1 & -1 & 0 & -1 & 0 & -1 \\
 0 & 0 & 1 & -1 & 0 & 0 & 0 \\
 0 & 0 & 0 & 1 & 1 & -1 & 0
 \end{bmatrix}
 \bullet
 \begin{array}{c}
 \boxed{\mathbf{V}} \\
 v_1 \\
 v_2 \\
 v_3 \\
 v_4 \\
 v_5 \\
 \boxed{v_6} \\
 v_7
 \end{array}
 =
 \begin{array}{c}
 0 \\
 0 \\
 0 \\
 0 \\
 0
 \end{array}
 \end{array}$$

Genome Annotations Contain Knowledge Gaps



Flux Balance Analysis



~~| | 1 | 2 | 3 | 4 | 5 | 6 | 7 |
|---|---|----|----|----|----|----|----|
| A | 1 | -1 | 0 | 0 | 0 | 0 | 0 |
| B | 0 | 1 | -1 | 0 | -1 | 0 | -1 |
| C | 0 | 0 | 1 | -1 | 0 | 0 | 0 |
| D | 0 | 0 | 0 | 1 | 1 | -1 | 0 |

 $\bullet$

V
v_1
v_2
v_3
v_4
v_5
v_6
v_7

 $=$

0
0
0
0
0
0

Model Auto-completion Optimization

Objective:

Minimize $\sum_{i=1}^r \left[z_{i,forward,not\ in\ model} + z_{i,reverse,reversible\ not\ in\ model} + 2z_{i,reverse,irreversible\ not\ in\ model} + z_{i,reverse,irreversible\ in\ model} \right]$

Penalizing reversibility adjustments

Penalizing addition of reactions to the model

Subject to:

Mass balance constraints:

		Reactions in model	Reactions not in model	
Compounds in model	N_{core}	N_{db}		$\begin{bmatrix} v_{core} \\ v_{db} \end{bmatrix} = 0$
Compounds not in model	0	N_{db}		

Use variable constraints:

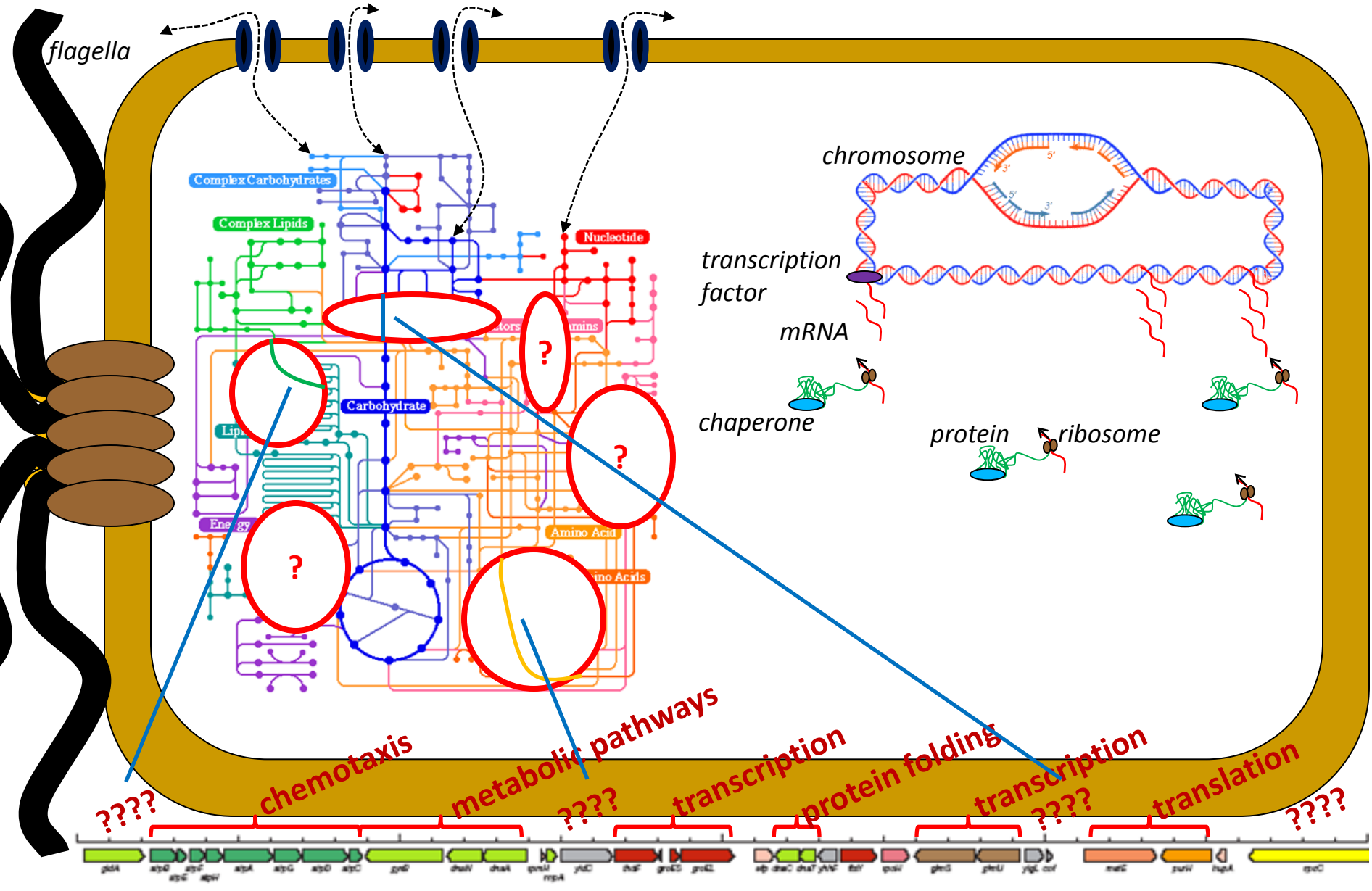
$$0 \leq v_{i,forward} \leq v_{Max} z_{i,forward}$$

$$0 \leq v_{i,reverse} \leq v_{Max} z_{i,reverse}$$

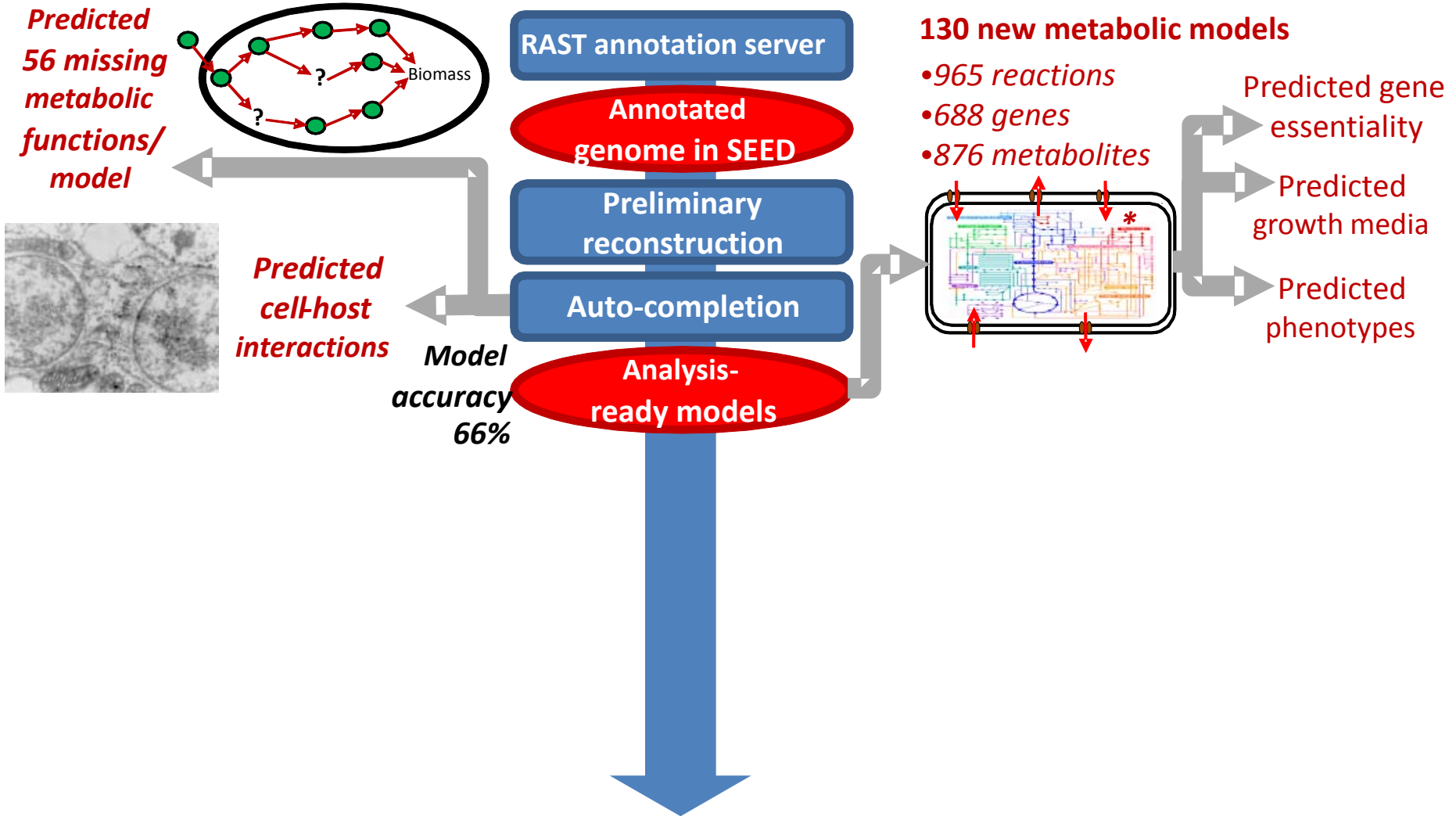
Forcing positive growth:

$$v_{biomass} > 0$$

Genome Annotation: the Subsystems Approach



Model SEED: Converting Annotated Genomes into Genome-scale Metabolic Models

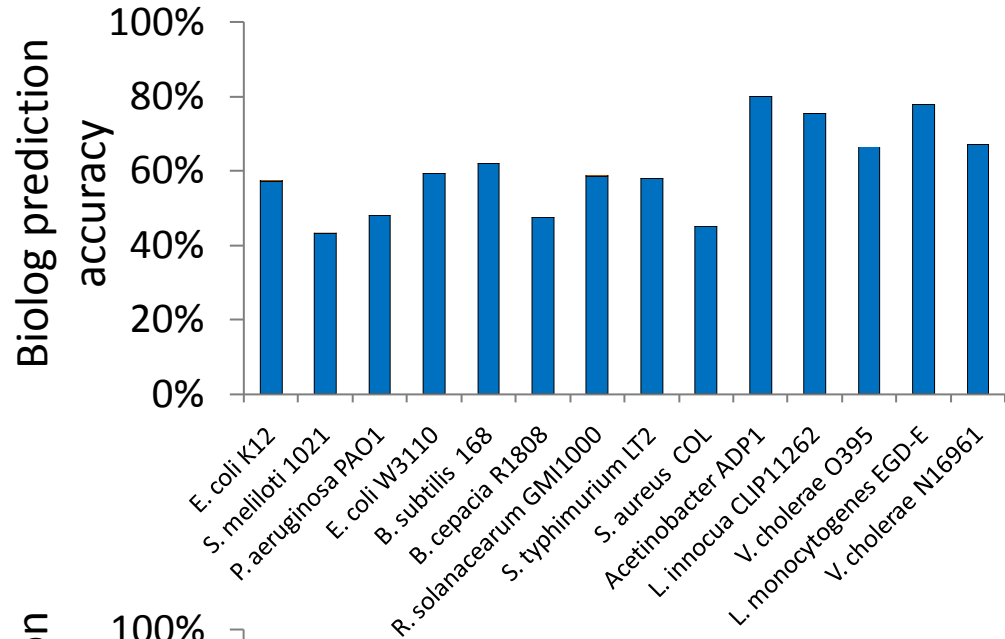


Accuracy Before Optimization

Biolog phenotype data

- SEED models were used to predict the output of 14 biolog phenotyping arrays

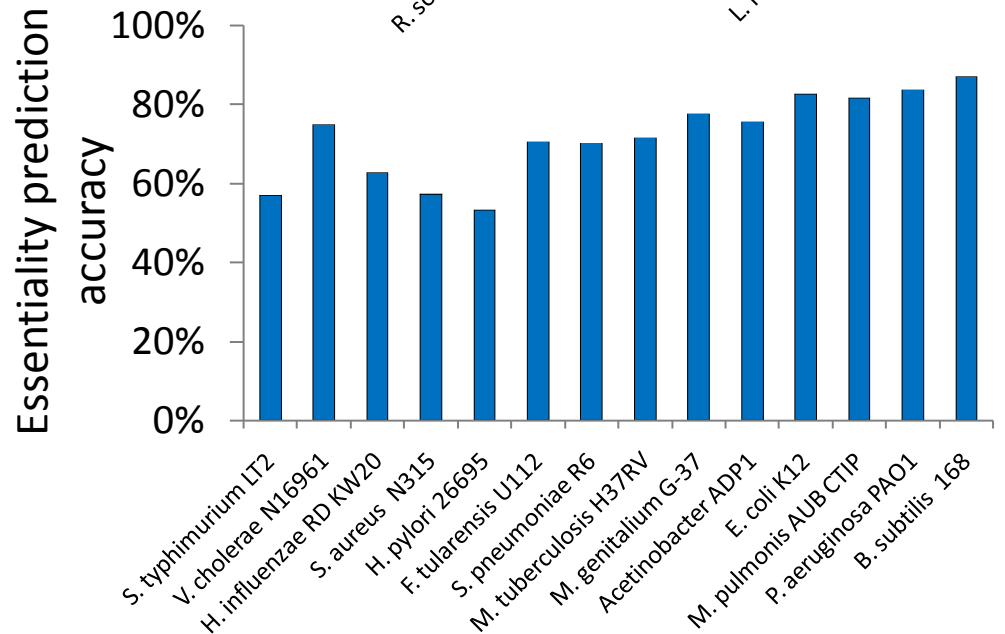
- Average accuracy: 60%



Essentiality data

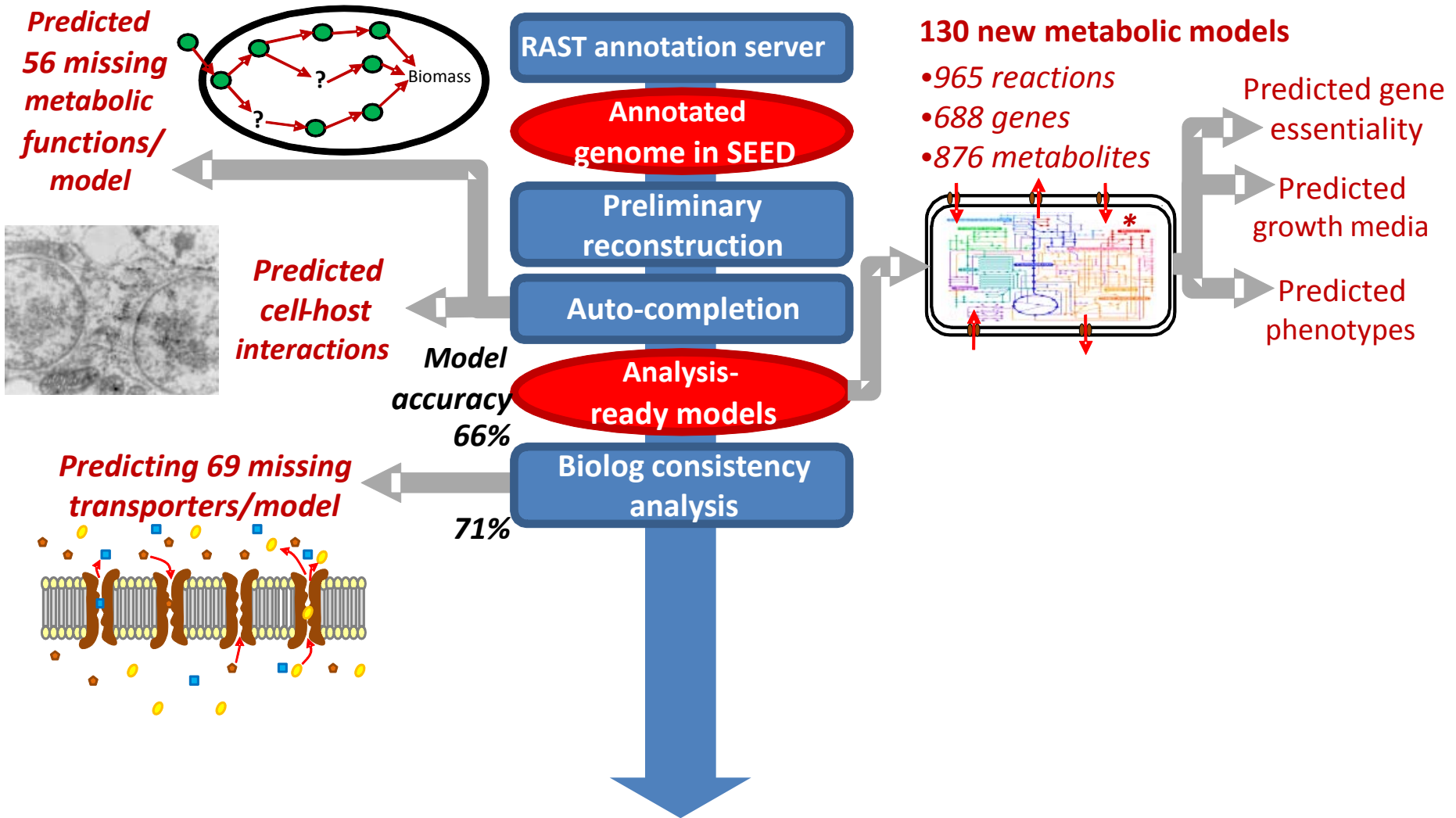
- SEED models were used to predict essential genes for 14 experimental gene essentiality datasets

- Average accuracy: 72%



Overall accuracy: 66%

Model SEED: Converting Annotated Genomes into Genome-scale Metabolic Models



Biolog Consistency Analysis

Biolog phenotype data

- Add transporters for Biolog nutrients if missing from models

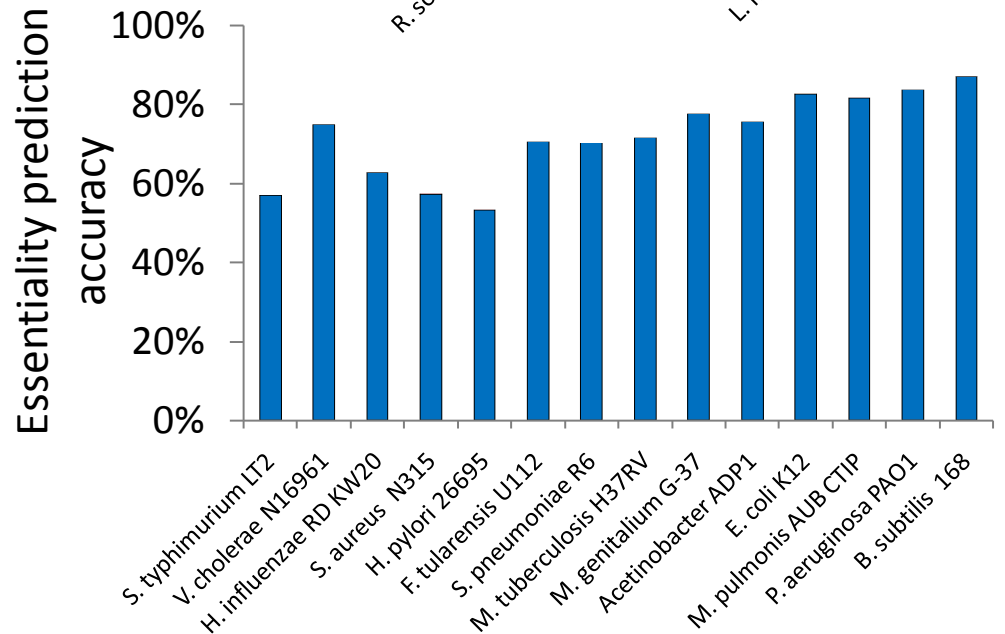
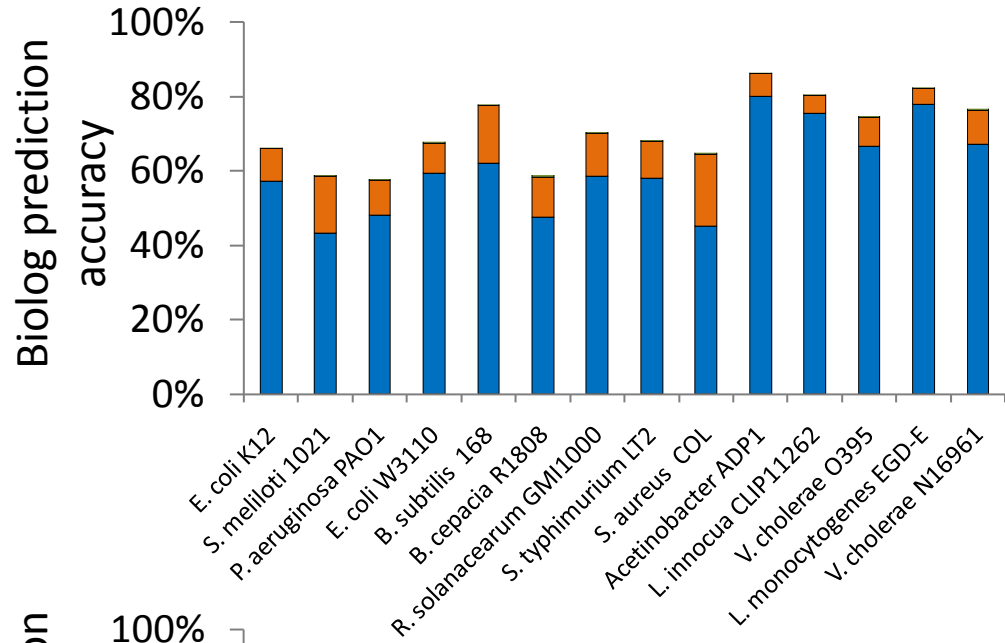
- 69 transporters added to each model on average

- Average accuracy: 70%

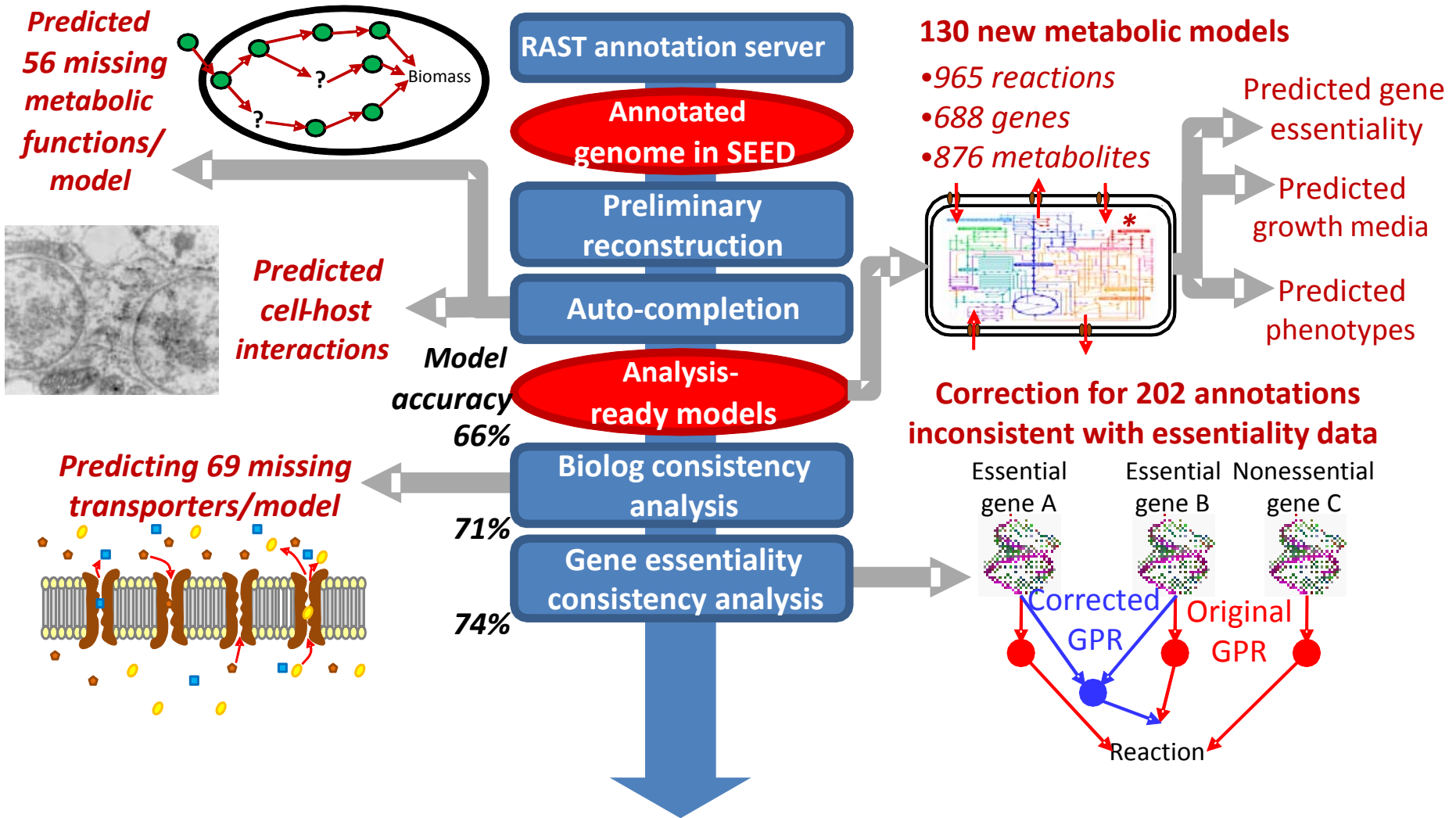
Essentiality data

- Accuracy unchanged: 72%

Overall accuracy: 71%



Model SEED: Converting Annotated Genomes into Genome-scale Metabolic Models



Annotation Consistency Analysis

Essentiality data

- Reconciling annotation inconsistent with essentiality data

Essential gene

Nonessential gene

Essential gene A

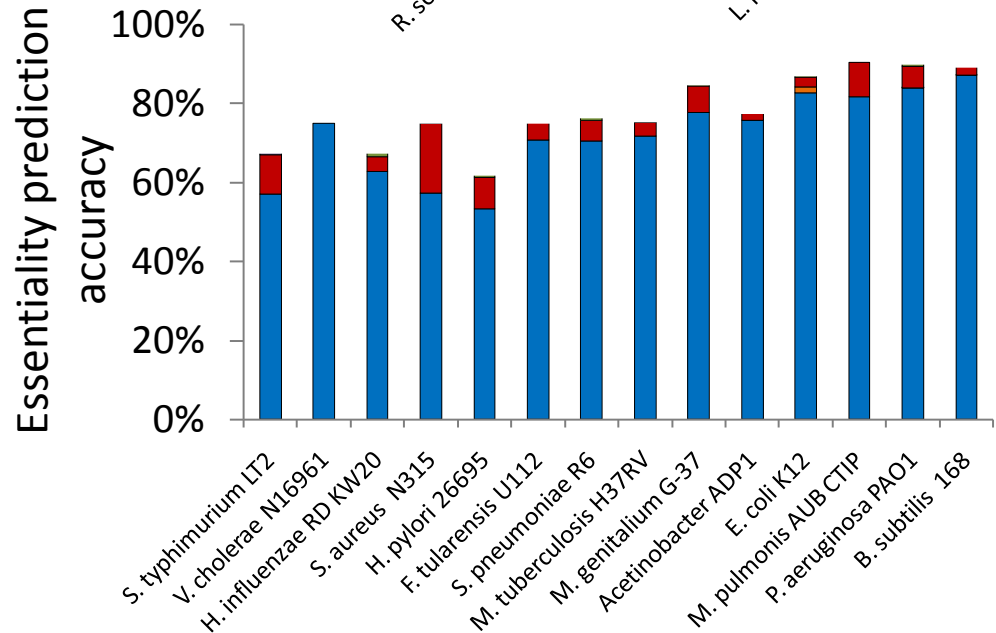
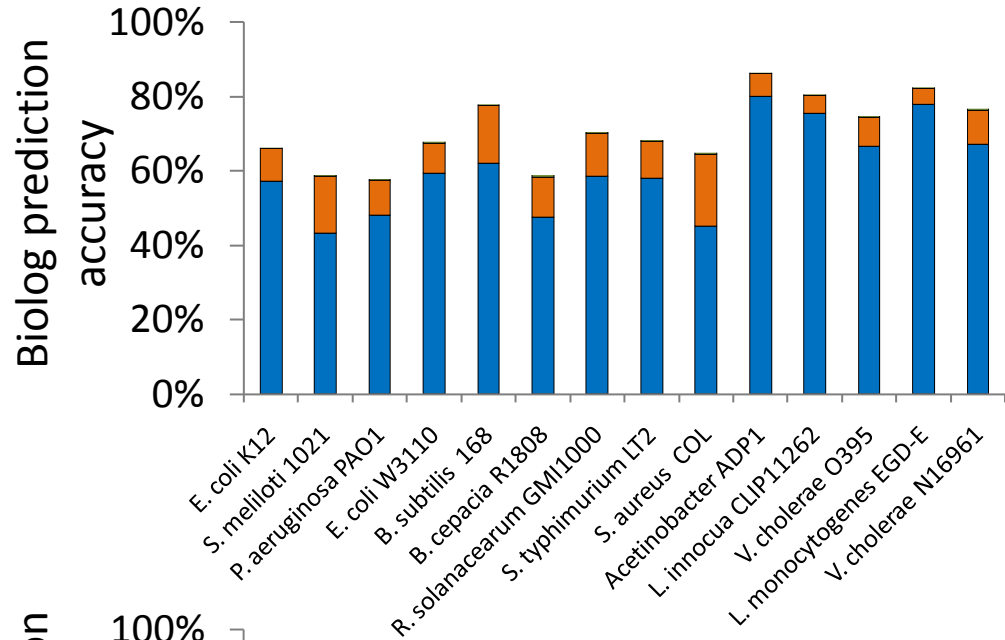
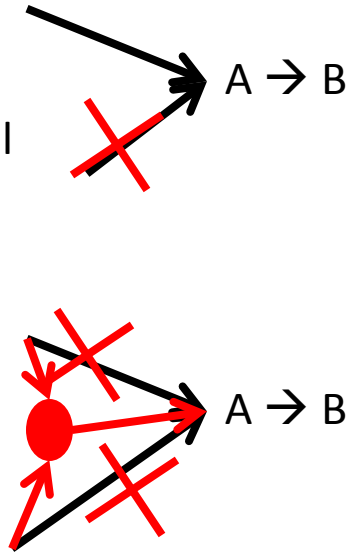
Essential gene B

- Accuracy 78%

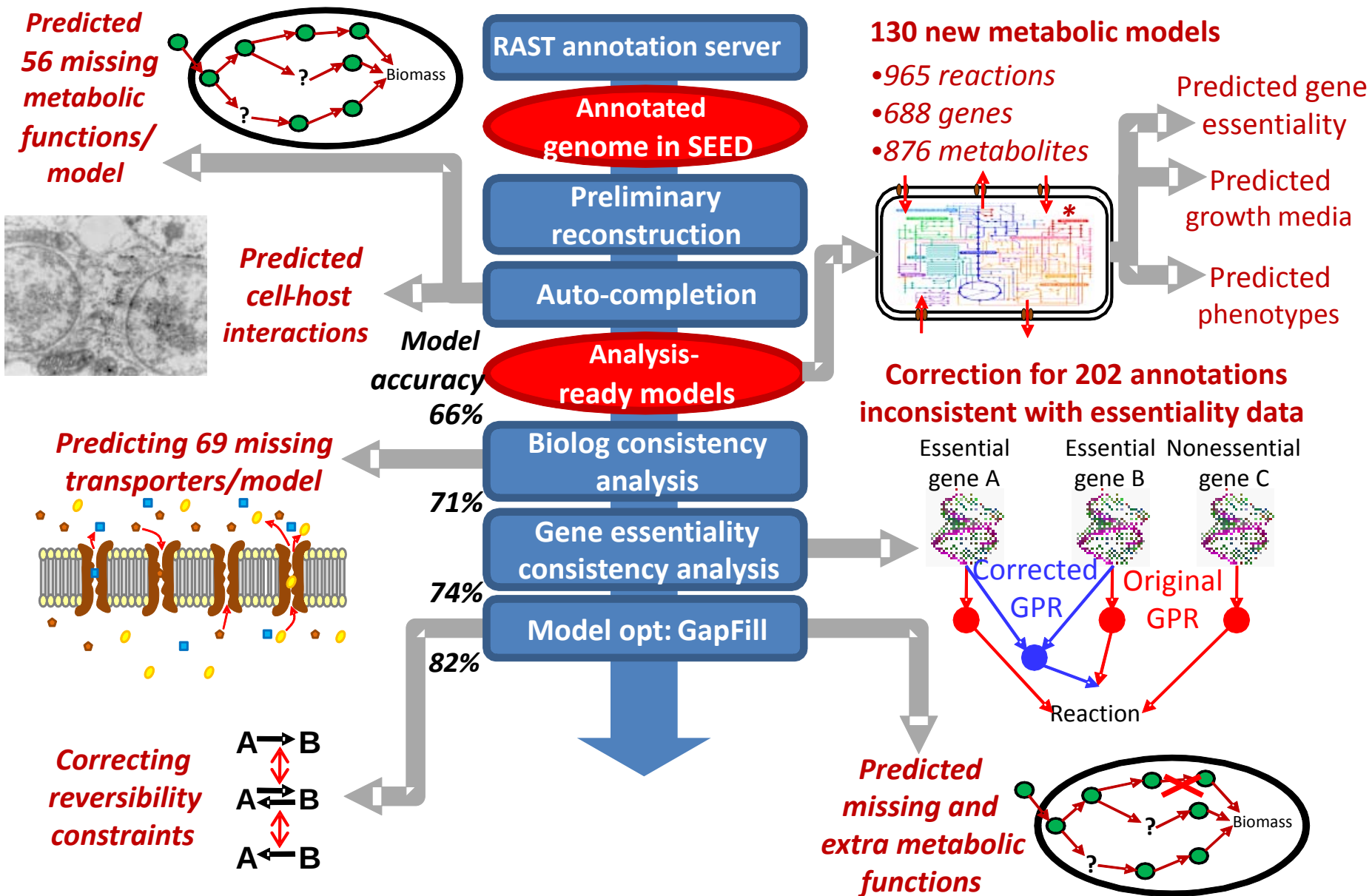
Biolog phenotype data

- Accuracy unchanged: 70%

Overall accuracy: 75%

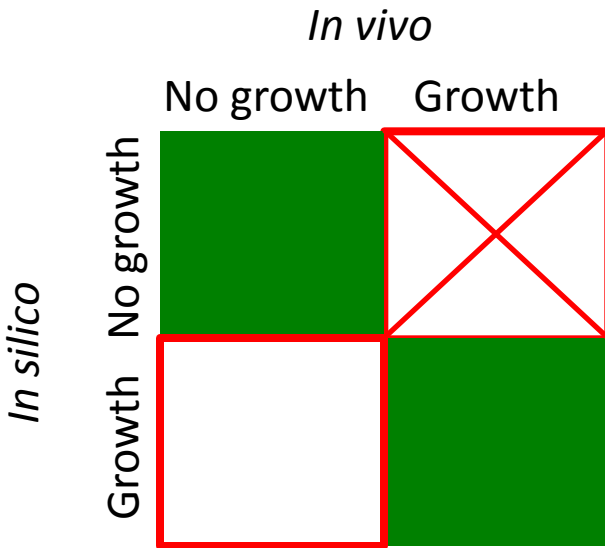


Model SEED: Converting Annotated Genomes into Genome-scale Metabolic Models



Model Optimization: Gap Filling

Additional gap filling:



- Fix false negative predictions by adding reactions to models

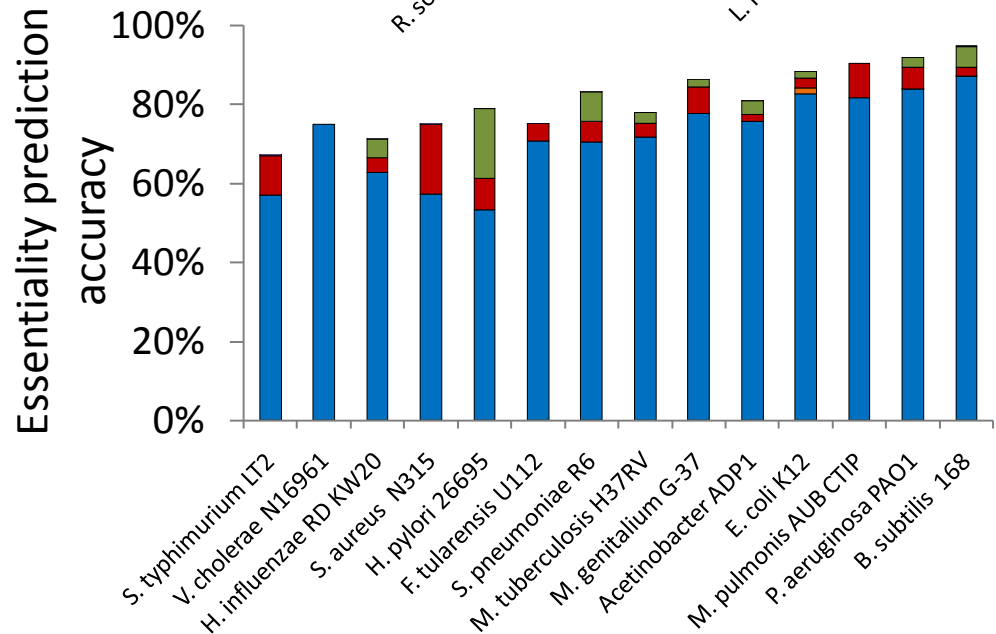
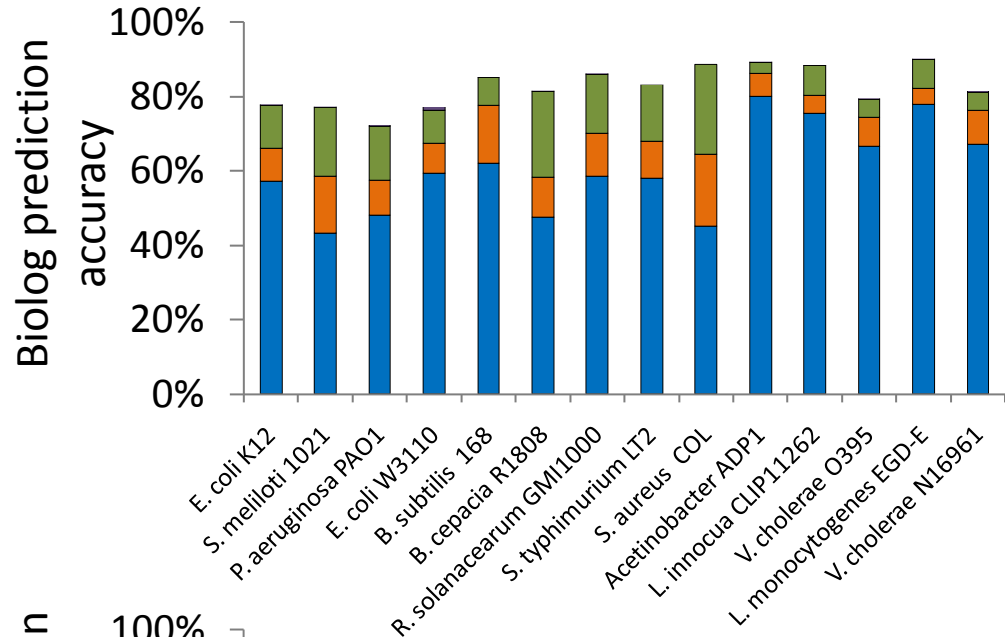
Biolog accuracy

- Average accuracy: 83%

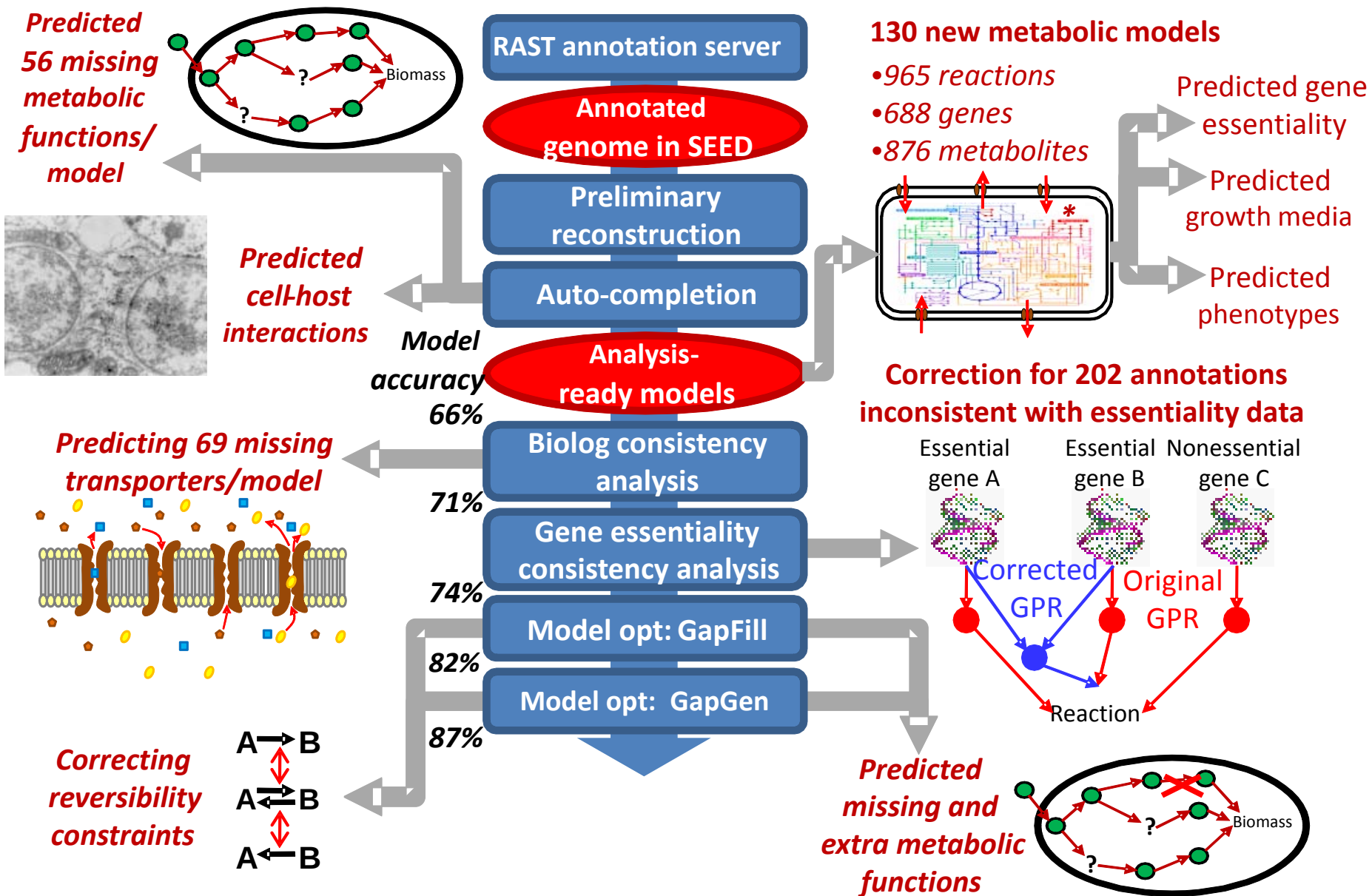
Essentiality accuracy

- Average accuracy: 81%

Overall accuracy: 82%



Model SEED: Converting Annotated Genomes into Genome-scale Metabolic Models



Model Optimization: Gap Generation

Additional gap filling:

		<i>In vivo</i>	
		No growth	Growth
<i>In silico</i>	No growth		
	Growth		

- Fix false positive predictions by removing reactions from models

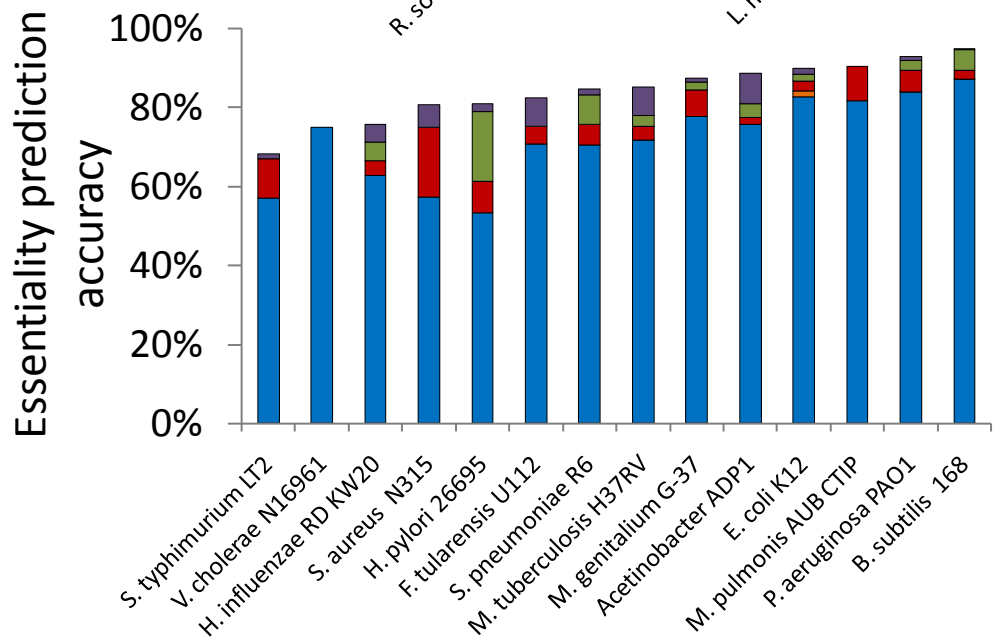
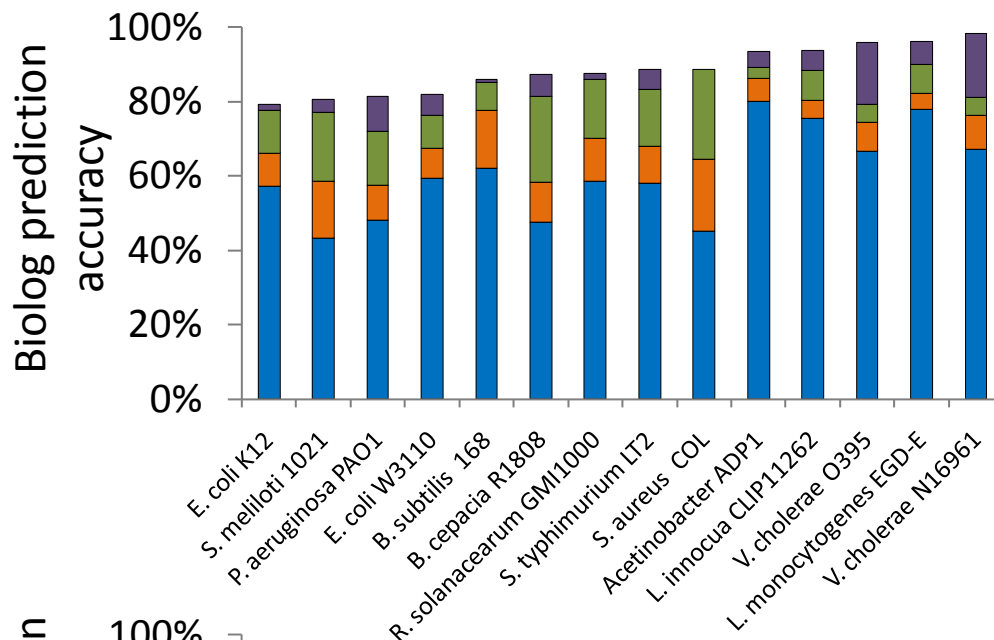
Biolog accuracy

- Average accuracy: 88%

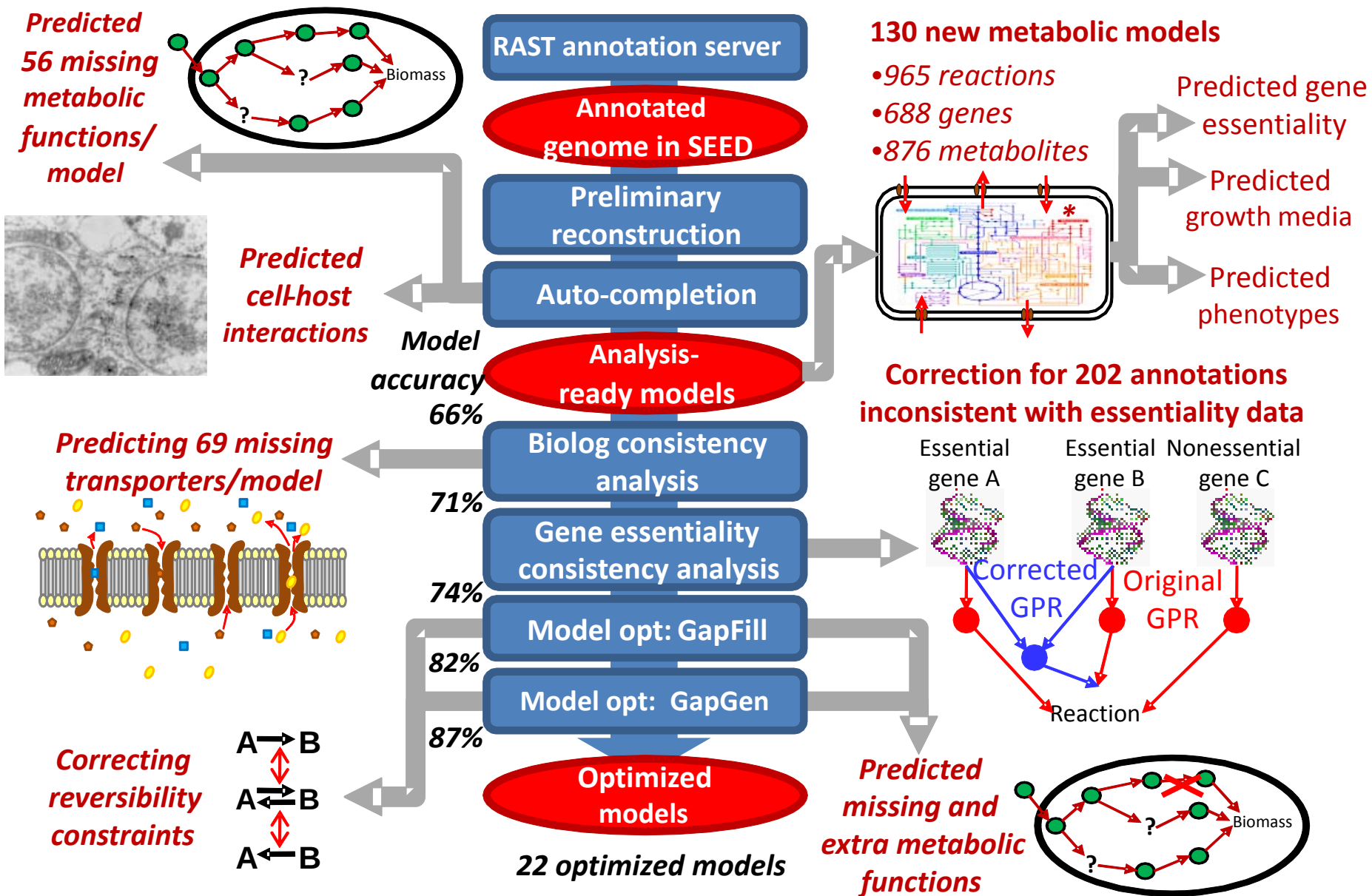
Essentiality accuracy

- Average accuracy: 85%

Overall accuracy: 87%



Model SEED: Converting Annotated Genomes into Genome-scale Metabolic Models



Seed Models vs Published Models

•Single-genome Seed models compare favorably with published single genome models

Organism name	Published model	Published reactions	SEED Reactions	Published genes	SEED genes
<i>Acinetobacter</i>	iAbaylyiv4	868	1196	775	785
<i>B. subtilis</i>	iYO844	1020	1463	844	1041
<i>C. acetobutylicum</i>	iJL432	502	989	432	721
<i>E. coli</i>	iAF1260	2013	1529	1261	1083
<i>G. sulfurreducens</i>	iRM588	523	721	588	468
<i>H. influenzae</i>	iCS400	461	969	400	575
<i>H. pylori</i>	iIT341	476	731	341	421
<i>L. plantarum</i>	iBT721	643	908	721	699
<i>L. lactis</i>	iAO358	621	965	358	646
<i>M. succiniciproducens</i>	iTK425	686	1048	425	659
<i>M. tuberculosis</i>	iNJ661	939	1021	661	728
<i>M. genitalium</i>	iPS189	264	294	189	214
<i>N. meningitidis</i>	iGB555	496	903	555	560
<i>P. gingivalis</i>	iVM679	679	744	0*	399
<i>P. aeruginosa</i>	iMO1056	883	1386	1056	1094
<i>P. putida</i>	iNJ746	950	1261	746	1053
<i>R. etli</i>	iOR363	387	1264	363	1242
<i>S. aureus</i>	iSB619	641	1115	619	770
<i>S. coelicolor</i>	iIB700	700	1159	700	987

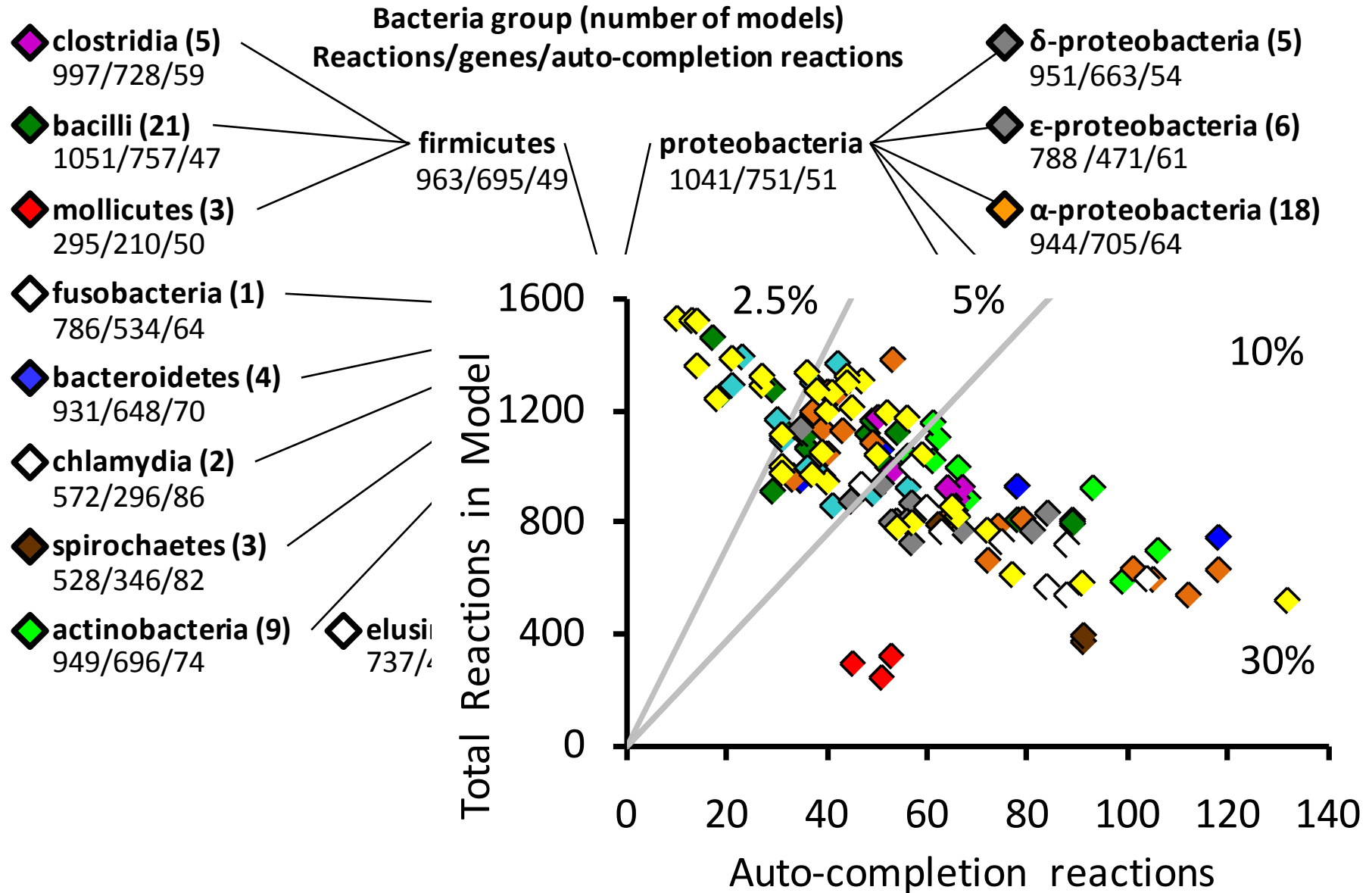
Assessing Subsystem Annotations From Auto-completion

- We identify how *complete* the annotations are for each of the Seed subsystems by calculating the following ratio:

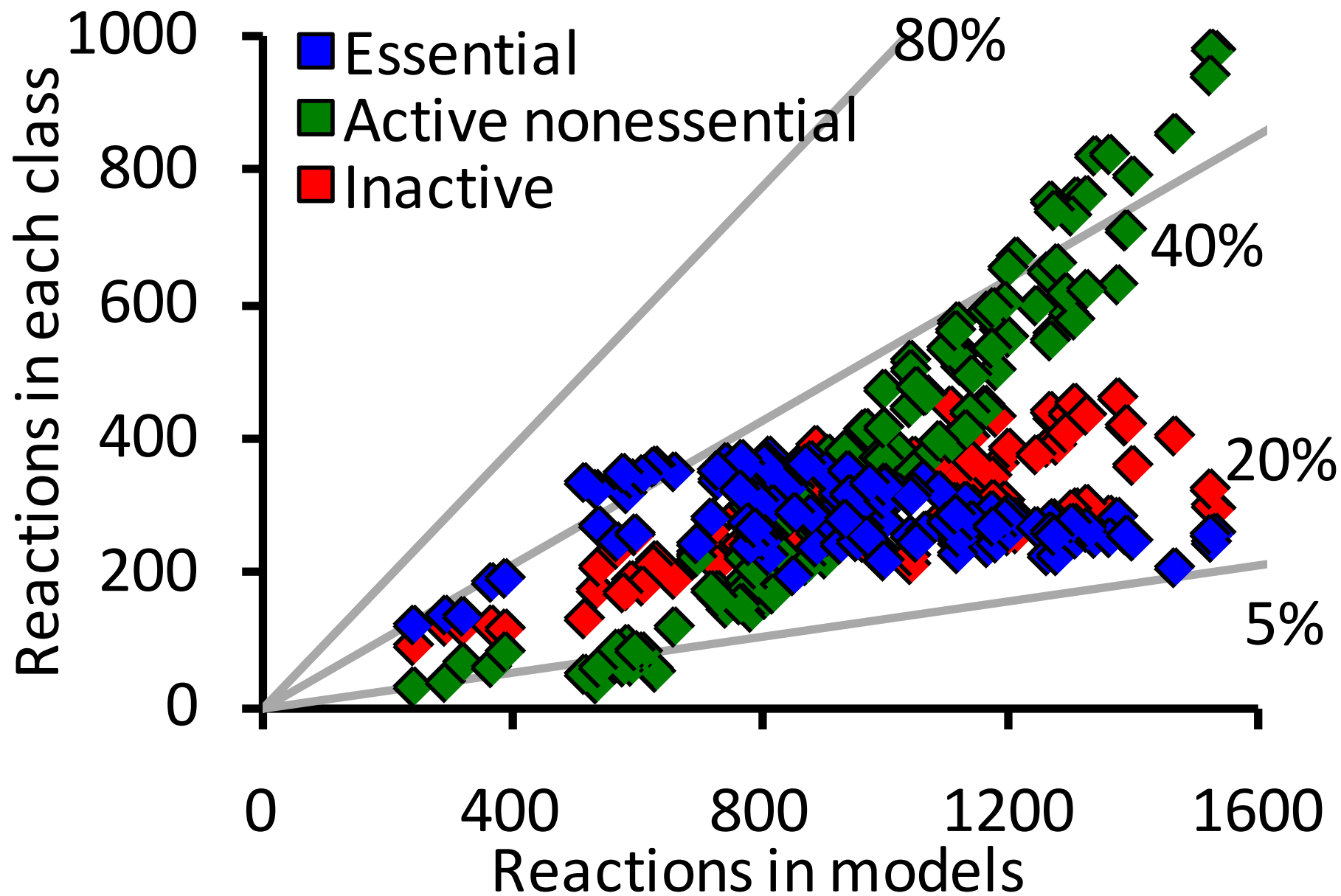
$$\frac{\text{auto-completion reactions in subsystem}}{\text{total reactions in subsystem}} = \text{Fraction of subsystem reactions with missing genes}$$

- Highest scoring subsystems:
- Cell Wall and Capsule Biosynthesis (15%)*
 - 21 reactions per model added during auto-completion
 - LOS Core Oligosaccharide Biosynthesis* (Gram negative)
 - Teichoic and Lipoteichoic Acids Biosynthesis* (Gram positive)
 - KDO2-Lipid A Biosynthesis*
- Cofactors, Vitamins, and Prosthetic Group Biosynthesis (5%)*
 - 10 reaction per model added during auto-completion
 - Ubiquinone Biosynthesis*
 - Menaquinone and Phylloquinone Biosynthesis*
 - Thiamin Biosynthesis*
- Six subsystems account for 31/56 reactions added to each model during the auto-completion process

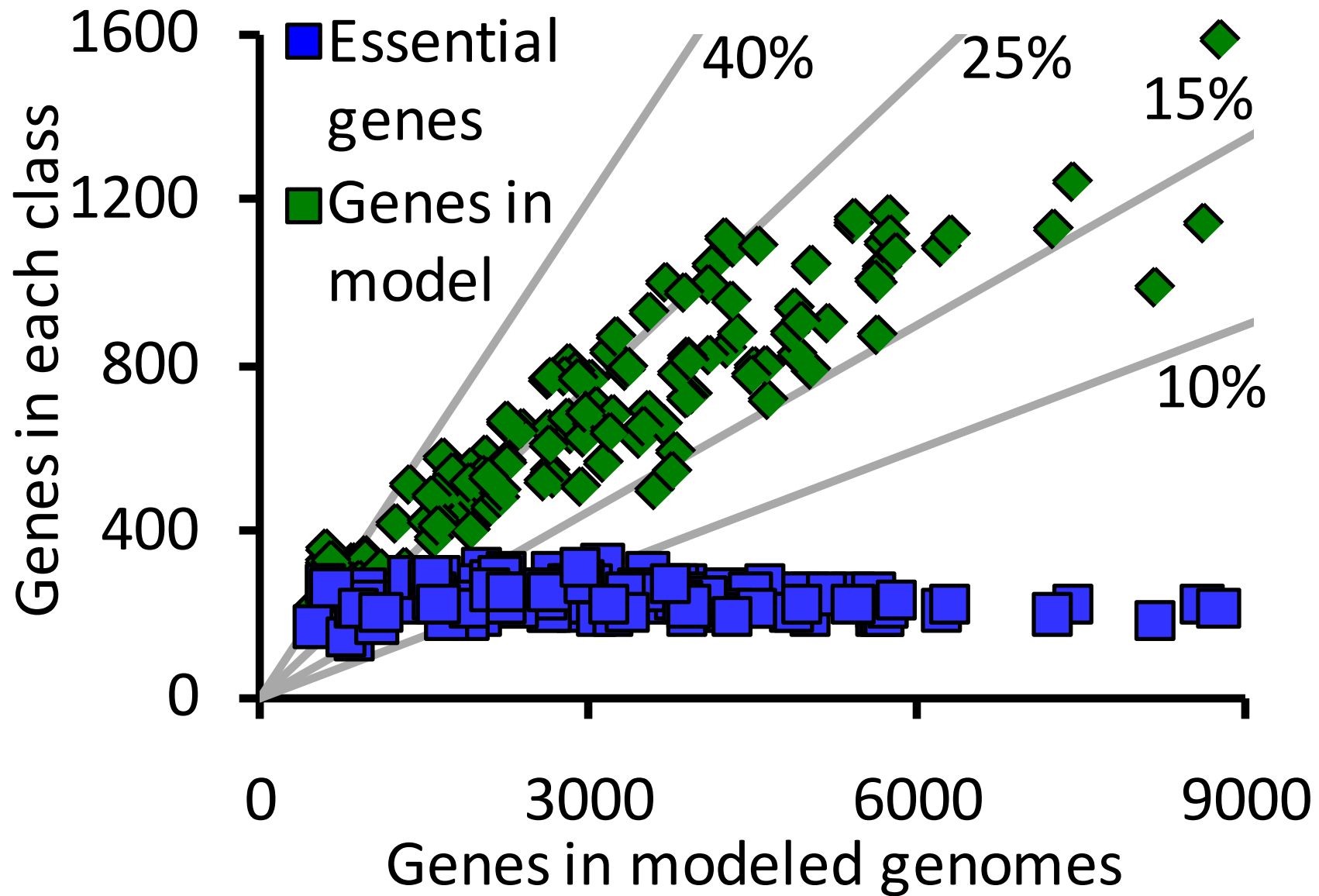
Model statistics across the phylogenetic tree



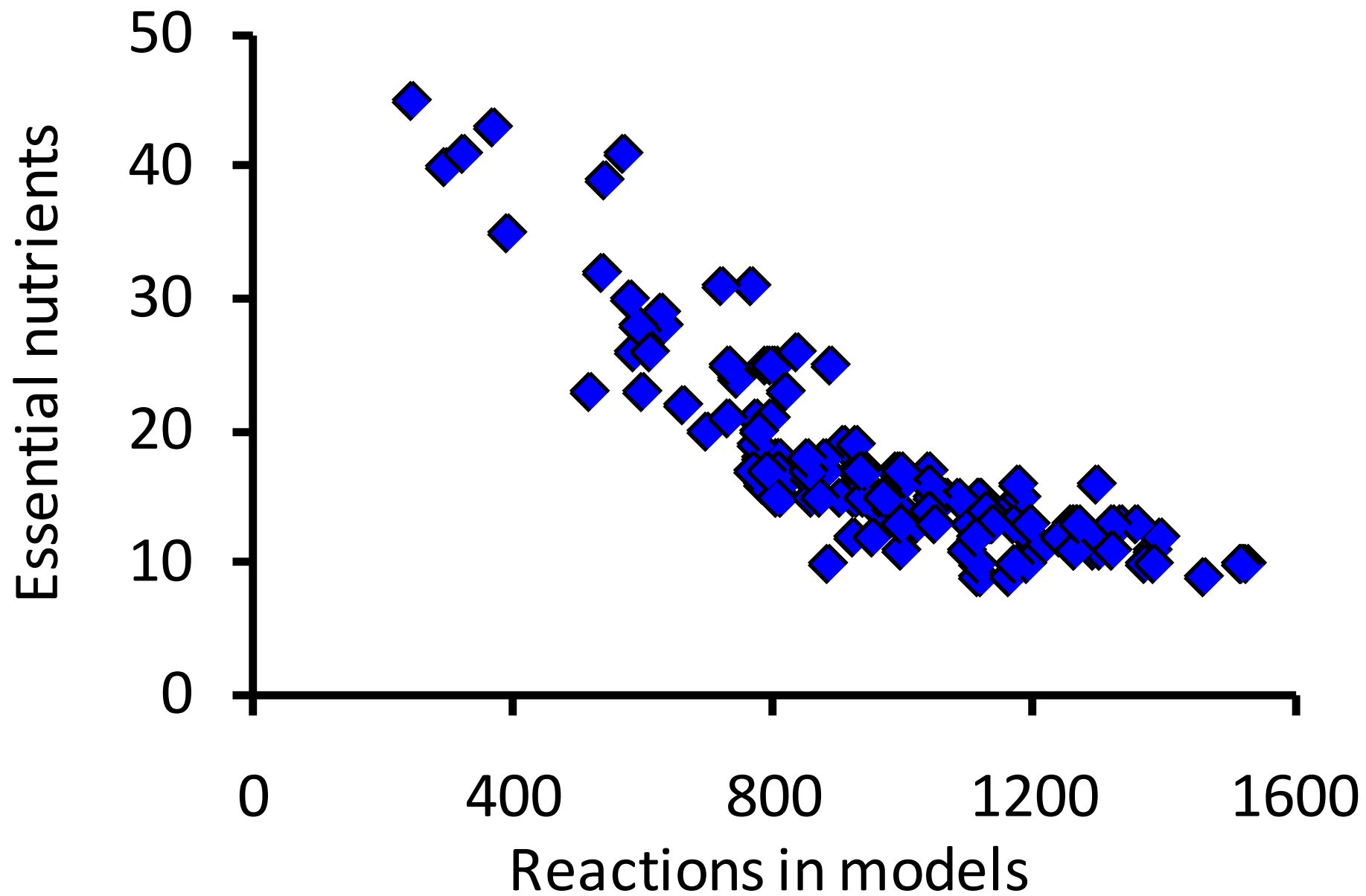
Reaction Activity Across All Models



Essential Genes Across All Models



Essential Nutrients Across All Models



Acknowledgements

ANL/U. Chicago Team

- Robert Olson
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- Olga Zagnitzko
- Svetlana Gerdes



Fellowship for
Interpretation of
Genomes

Hope College Team

- Aaron Best
- Matt DeJongh
- Nathan Tintle
- Hope college students



National Institute of Allergy and Infectious Diseases
National Institutes of Health